

# International Journal of Bio-resource and Stress Management

Print ISSN 0976-3988

IJBSM 2021, 12(6):655-662

Online ISSN 0976-4038

Crossref

December 2021

Article AR2209b

Research Article

Natural Resource Management

# Effect of Pre-sowing Treatments on Vegetative Growth and Production of Graftable Rootstocks in Mango

Sanjeev Kumar Banyal<sup>1\*</sup>, Shiv Kumar Shivandu<sup>2</sup> and Uday Raj Patial<sup>3</sup>

<sup>1</sup>Dept. of Fruit Science, College of Horticulture and Forestry, Dr. Y S Parmar University of Horticulture and Forestry, Neri, Hamirpur, H.P. (177 001), India

<sup>2</sup>Dept. of Fruit Science, Dr. Y S Parmar University of Horticulture and Forestry, Solan HP (173 230), India <sup>3</sup>Dept. of Fruit Science, PAU Ludhian, Punjab (141 004), India



Sanjeev Kumar Banyal e-mail: skbanyal@gmail.com

Citation: Banyal et al., 2021. Effect of Presowing Treatments on Vegetative Growth and Production of Graftable Rootstocks in Mango. International Journal of Bio-resource and Stress Management 2021, 12(6), 655-662. HTTPS://DOI. ORG/10.23910/1.2021.2209b.

Copyright: © 2021 Samant et al. This is an open access article that permits unrestricted use, distribution and reproduction in any medium after the author(s) and source are credited.

Data Availability Statement: Legal restrictions are imposed on the public sharing of raw data. However, authors have full right to transfer or share the data in raw form upon request subject to either meeting the conditions of the original consents and the original research study. Further, access of data needs to meet whether the user complies with the ethical and legal obligations as data controllers to allow for secondary use of the data outside of the original study.

Conflict of interests: The authors have declared that no conflict of interest exists.

#### Abstract

The present investigations "Studies on the effect of pre-sowing treatment and time of wedge grafting on production of saleable grafts in mango" were carried out in the nursery block of Fruit Science Department. The study was conducted using wild mangoes seeds. The seeds were divided in two equal halves, one half of seeds were sown with intact seed coat (K<sub>1</sub>) while, in other half the mango kernels were extracted (K<sub>2</sub>), both type of seeds were then soaked in 6 different chemicals of varying concentrations viz.  $C_1$ -  $KNO_3$  @ 0.5%,  $C_2$ -  $KNO_3$  @ 1%,  $C_3$ -  $KNO_3$  @ 1.5%, C<sub>4</sub>- BA @ 50 ppm, C<sub>5</sub>- BA @ 100 ppm, C<sub>6</sub>- GA<sub>3</sub> @ 100 ppm, C<sub>7</sub>- GA<sub>3</sub> @ 200 ppm, C<sub>8</sub>beejaaumrit @ 2%,  $C_9$  - beejaaumrit @ 3%,  $C_{10}$  - panchgavya @ 2%,  $C_{11}$  - panchgavya @ 3% for 12 hours and  $C_{12}$ - Control. The germination and vegetative parameters were observed. The results of study revealed that the extracted kernels presoaked with GA<sub>3</sub> @ 100 ppm produced earliest germination, 50 % germination, high rate of germination, germination percentage, number of leaves, maximum leaf area, highest seedling vigour index, while, on pre-soaking extracted kernel with KNO, @ 0.5% produced maximum seedling diameter. The Seedling raised from extracted kernels pre-soaked in KNO<sub>2</sub> @ 0.5% (C, K<sub>2</sub>) attained graftable diameter within 210 days of sowing. The present investigation was conducted during 2019-2020 at the nursery block of College of Horticulture and Forestry, Neri Hamirpur (H.P) to study the effect of pre-sowing treatments on germination, growth and production of graft-able seedlings.

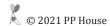
**Keywords:** Pre-sowing treatments, seedling emergence, mango

## 1. Introduction

King of fruits, "mango" is one of the most favoured fruit of every Indian household (Tharanathan et al., 2006; Paulmurugan, 2021) Origin can be traced back to Indo-Burma region belonging to anacardiaceae family having chromosome number 2n= 40 (Singh et al., 2016). Being perennial it is slow growing and take lot of time to bear fruits, however, taking into consideration the growing demand for the mango cultivation the more amount of planting material is required. Mango being a cross pollinated crop represent a wide variation in the plants propagated through seeds (Schnell et al., 2006). So, commercial mango is propagated through vegetative propagation. Rootstock is the base of successful graft union as it influences the vigor, life of tree, yield and its quality. It becomes mandatory to have healthy and strong rootstock for successfully grafted

**Article History** 

RECEIVED on 23rd March 2021 RECEIVED in revised form on 20th October 2021 ACCEPTED in final form on 16th December 2021



plants. In the last 20 years, India has witnessed an increase in area under mango cultivation as the demand for fresh fruits has taken a rise in domestic as well as global market. Being highly cross-pollinated in nature there is humongous variation in plants even if they are grown from the fruits of the same plant. Thus, it becomes highly necessary to preserve and maintain the desirable characters of each variety by propagation through vegetative methods.

Mango seed being recalcitrant (Berjak and Pammenter, 2004; Umarani et al., 2015) (12-50% germination after a month) and its availability is being restricted to drier part (May -July) of the year makes it equally important for the grower to utilize that time for growing stock. However, on sowing of mangos stones it was found that they have lower germination and survival rate. On using the pre-soaked mango seeds in GA (gibberellic acid) and KNO<sub>3</sub> (Potassium Nitrate) exhibited magnificent effect on germination and vegetative growth of the seedlings so geminated. The hard seed coat present on the mango kernels act as a barrier in germination and is the major reason for delay in the germination. Rapid germination leads to quick establishment of the seedling which prevents the damage to seed on its critical phase of growth (Harris et al., 1999). Many other factors such as the method of sowing and orientation of seed at the time of sowingare also responsible for the growth of seedlings (Ross and Harper, 1972). Sowing of seeds with stalk end up position leads to placement of seeds in most appropriate position resulting in early germination by stimulating metabolic process which initiate the rapid radical emergence (Bewley and Black, 2012; Rajjou et al., 2012). Major key factor having crucial impact on viability of mango stones is the lower content of moisture in mango seeds as the mango ages it can causes disruption of normal metabolic cycle of seed germination and hence, slows the germination process of seed (Patel et al., 2016a; Kozlowski and Pallardy, 1997). To access the effect of removal of seed coat for extraction of kernels and soaking of seeds in chemical with objective of early germination, improving germination percentage for the rapid production of healthy root stocks to fulfill the market demand and to know the best set of practices in order to produce the high quality mango planting material. This study is focused at to gather information regarding the mango seed germination and development of seedling in order to facilitate the production of rootstocks for a successful orchard establishment.

### 2. Materials and Methods

The present investigation was conducted during 2019–2020 at the nursery block of College of Horticulture and Forestry, Neri Hamirpur (H.P) the experiment was carried out using Randomized block design having 24 treatments combination having four replications. The 24 different combinations were performed by the use of 2 type of seeds (Intact stones and Extracted mango kernels) and 12 treatments for pre-soaking with chemicals namely, GA<sub>3</sub> @ 100 ppm and 200 ppm, KNO<sub>3</sub> @ 0.5%, 1% and 1.5%, benzyl adenine @ 50 ppm and 100 ppm, beejaaumrit @ 2% and 3 %, panchgavya @ 2% and 3% and control. Wild mango fruits were collected from the nearby villages and stones were extracted. The seeds were washed thoroughly to remove pulp adhering to the stones. Whole lot of mango stones were immersed in water bucket and the stones which settled at bottom were selected for the experiment further while, the stones which were floating were rejected. The mango stone were shade dried. Then the stones were divided in 2 equal number one half was use with the intact seed coat while in other half was used for extracting kernel. After that the intact stones and extracted kernels were soaked simultaneously in the abovementioned chemicals for the time period of 12 hours. The 24 treatments were replicated thrice having 100 seeds per replication and were sown in nursery beds.

Germination parameters were recorded and calculated:

Germination percentage=(number of seeds germinated )/ (Total number of seeds sown)×100

Rate of germination= (Number of Seed Germinated till 30 Days of sowing)/(Total Number of Seeds Sown)×100

The seedling vigour index on growth basis was calculated using the seedling height multiplied by germination percentage.

Percent survival=(Number of seedlings survived )/(Total number ofseedlings germinated)×100

Observations were recorded from all seedlings and average value was calculated and was represented. The initiation of germination was found within 8 days after sowing of extracted kernels while, sown stones required time of 18 days. Observation related to germination of seedling were recorded on daily basis while, those pertaining to vegetative parameters were recorded after interval of 30 days. The data so recorded was analyzed using the statistically procedure used by Gomez and Gomez (1984).

### 3. Results and Discussion

Germination and growth of mango seedlings was found significantly influenced by the use of pre-sowing treatments given to mango seeds.

# 3.1. Effect on germination

The data (Table 1) revealed that on soaking extracted kernel with GA<sub>2</sub> @ 100 ppm was found best to initiate the germination within 8.47 days while 50% germination was completed within the time of 13.65 days. Highest (90.38%) germination percentage and rate of germination (54.00%) was recorded in extracted kernels treated with GA, @100 ppm. This difference in germination parameters could be attribute to the presence of a hard seed coat outside the kernel which act as a barrier in uptake of water (Mwaurah et al., 2020), so the removal outer covering which increase the water absorption and cause hydrolysis of carbohydrate present in cotyledons leading to better germination (Kaur et al., 1998; Paleg 1960; Sanchez

Chemicals	Days	for initiation of	germination	Days for 50% germination			
	K <sub>1</sub> (Intac mango sto			K₁ (Intact mango stor		ed Mean	
C <sub>1</sub> (KNO <sub>3</sub> @ 0.5%)	30.72	9.87	19.63	42.65	13.72	28.18	
C <sub>2</sub> (KNO <sub>3</sub> @ 1%)	34.75	9.52	22.14	46.93	16.17	31.55	
C <sub>3</sub> (KNO <sub>3</sub> @ 1.5%)	35.58	9.46	22.52	41.65	16.00	28.83	
C <sub>4</sub> (BA @ 50 ppm)	32.48	9.97	21.23	49.45	15.33	32.39	
C <sub>5</sub> (BA @ 100 ppm)	38.23	12.8	1 25.52	48.56	17.60	33.08	
C <sub>6</sub> (GA <sub>3</sub> @ 100 ppm)	22.20	8.47	15.33	36.76	13.65	25.20	
C <sub>7</sub> (GA <sub>3</sub> @ 200 ppm)	25.93	8.54	17.90	39.64	15.45	27.54	
C <sub>8</sub> (Beejaumrit @ 2%)	40.85	17.6	7 29.26	51.23	23.48	37.36	
C <sub>9</sub> (Beejaumrit @ 3%)	41.94	16.0	1 28.98	56.34	23.75	40.05	
C <sub>10</sub> (Panchgavya @ 2%)	34.15	13.54	4 23.84	45.76	17.91	31.84	
C <sub>11</sub> (Panchgavya@3%)	35.37	11.4	7 23.42	43.21	14.98	29.09	
C <sub>12</sub> (Control)	42.27	22.60	6 32.47	58.17	35.67	46.92	
Mean	34.54	12.49	9	46.70	18.64		
SEm±		K 1	.49 .20 .70		C 0.38 K 0.92 C×K 1.30	2	
CD ( <i>p</i> =0.05)		C 0 K 2	.95 .31 .31		C 1.76 K 4.20 C×K NS	; )	
Chemicals	Germ	ination percenta		Rate	e of germination		
	K <sub>1</sub> (Intact mango stone)	K <sub>2</sub> (Extracted Kernel)	Mean	K <sub>1</sub> (Intact mango stone)	K <sub>2</sub> (Extracted kernel)	Mean	
C <sub>1</sub> (KNO <sub>3</sub> @ 0.5%)	61.33 (51.59)	88.74 (70.40)	75.04 (60.99)	22.00 (27.95)	43.33 (41.09)	32.67 (34.52)	
C <sub>2</sub> (KNO <sub>3</sub> @ 1%)	60.00 (50.78)	72.23 (58.19)	66.12 (54.49)	20.67 (26.95)	42.00 (40.37)	31.33 (33.66)	
C <sub>3</sub> (KNO <sub>3</sub> @ 1.5%)	59.33 (50.49)	73.49 (59.09)	66.41 (54.79)	20.67 (26.81)	39.67 (39.02)	30.17 (32.92)	
C <sub>4</sub> (BA @ 50 ppm)	55.67 (50.36)	80.67 (63.90)	70.00 (57.13)	14.33 (22.20)	40.33 (39.39)	27.33 (30.80)	
C <sub>5</sub> (BA @ 100 ppm)	71.33 (48.26)	76.45 (60.97)	66.06 (54.61)	14.33 (22.20)	35.33 (36.40)	24.83 (29.30)	
C <sub>6</sub> (GA <sub>3</sub> @ 100 ppm)	64.67 (57.62)	90.38 (73.34)	80.86 (65.48)	24.00 (29.29)	54.00 (47.33)	39.00 (38.31)	
C <sub>7</sub> (GA <sub>3</sub> @ 200 ppm)	56.00 (53.52)	78.88 (57.42)	67.77 (55.47)	24.33 (29.54)	41.67 (40.14)	33.00 (34.84)	
C <sub>8</sub> (Beejaumrit @ 2%)	56.00 (48.43)	61.81 (51.92)	58.91 (50.18)	14.33 (22.22)	40.00 (39.21)	27.17 (30.72)	
C <sub>a</sub> (Beejaumrit @ 3%)	55.00 (47.86)	67.22 (55.22)	61.11 (51.54)	14.33 (22.14)	36.67 (37.23)	25.50 (29.69)	
C <sub>10</sub> (Panchgavya @ 2%)	57.00 (49.01)	75.71 (60.50)	66.36 (54.76)	17.00 (24.32)	35.33 (36.43)	26.17 (30.37)	
C <sub>11</sub> (Panchgavya@3%)	59.67 (50.57)	71.74 (57.92)	65.70 (54.24)	16.00 (23.32)	30.00 (33.17)	23.00 (28.25)	
C <sub>12</sub> (Control)	47.67 (43.64)	51.21 (45.67)	49.44 (44.66)	10.67 (19.02)	19.67 (26.13)	15.17 (22.58)	
Mean	58.92 (50.18)	73.38 (59.54)		17.72 (24.66)	38.17 (37.99)		
SEm±		C 1.07 K 2.62 C×K 3.70			C 0.39 K 0.94 C×K 1.33		
CD ( <i>p</i> =0.05)		C 1.76 K 4.32 C×K 6.11			C 1.59 K 6.18 C×K NS		

et al., 1967).  $GA_3$  at lower doses were found responsible for enhancing the enzymatic processes and lead to suppression of inhibitors. The early germination in extracted kernels might be the result of presence of no active barrier in germination which caused the easy absorption of the chemicals and  $GA_3$  caused stimulation of the radical and plumule to initiate germination, 50% germination, germination percentage and rate of germination (Kumar, 2007). The above results were found in conformity with the results of Aatla and Srihari (2013) in mango, and Patel et al. (2016a).

#### 3.2. Vegetative parameters

Maximum diameter and least number of days to obtain maximum seedling girth were observed in seedlings raised from extracted kernels pre-soaked with KNO<sub>3</sub> @ 0.5%. This might be due the initiation of early accumulation of the food material in the seedling tissues which helped in obtaining the maximum seedling diameter. The above results were found in compliance with the reports provided by Aatla and Srihari (2013), Patel et al. (2016b), Kumar et al. (2008), Padma and Reddy (1997) in mango (Table 2).

Whereas, On using extracted kernels pre-soaked in  $GA_3$  @ 100 ppm for sowing in nursery beds and the seedlings so raised were found having maximum seedling height which might be due to the stimulatory action of  $GA_3$  on the axillary and apical meristematic cells causing rapid division and multiplication of

the cells (Mauseth, 1976) leading to increase in seedling height also the removal of outer hard seed coat also aids in the better absorption of the chemicals and lead to early activation of amylase enzyme leading to conversion of the starch present in the cotyledons to sugars hence, enhancing the seedling height. However, the influence if GA, on the intercalary meristem was also found adding up to increase in the seedling height (Dohono and Walker, 1957; Gupta and Chakrabarty, 2013) which might have resulted in the production of thin stems because the plants were found more oriented toward the vertical growth rather being focused on lateral growth as the KNO<sub>3</sub> @ 0.5% treated seedlings did. Whereas, the other reason for better growth of seedling might be presence of higher number of leaves per plant which leads to production of higher photosynthetic assimilate to be utilized by plant for growth (Loach, 1970) (Table 3 and Figure 1).

Maximum number of leaves per plant might be due to the movement of gibberellins in to shoot apex which increases cell division and cell growth apparently leading to increase in development of young leaves (Salisbury and Ross, 1992) which lead to producing higher number of leaves per plant at much higher rate (Brijwal and Kumar, 2013) in comparison to seedlings under other treatments. The similar results were reported by, Kumar et al. (2008), Shaban (2010), Aatla and Srihari (2013), Sharaf (2016) in Rangpur lime and Choudhari and Chakrawar (1981) in citrus.

Table 2: Effect of pre-soaking treatments on vegetative growth of seedlings

Chemicals	Height			Diameter			
	K <sub>1</sub> (Intact mango stone)	K <sub>2</sub> (Extracted Kernel)	Mean	K <sub>1</sub> (Intact mango stone)	K <sub>2</sub> (Extracted kernel)	Mean	
C <sub>1</sub> (KNO <sub>3</sub> @ 0.5%)	26.26	28.28	27.27	7.26	8.24	7.75	
C <sub>2</sub> (KNO <sub>3</sub> @ 1%)	26.36	28.57	27.47	5.94	7.22	6.58	
C <sub>3</sub> (KNO <sub>3</sub> @ 1.5%)	24.65	29.45	27.05	5.76	7.29	6.53	
C <sub>4</sub> (BA @ 50 ppm)	24.37	28.86	26.61	5.87	6.84	6.36	
C <sub>5</sub> (BA @ 100 ppm)	25.64	26.99	26.32	6.37	7.25	6.81	
C <sub>6</sub> (GA <sub>3</sub> @ 100 ppm)	27.25	39.15	33.20	6.49	7.82	7.15	
C <sub>7</sub> (GA <sub>3</sub> @ 200 ppm)	26.97	35.76	31.37	6.14	7.53	6.83	
C <sub>8</sub> (Beejaumrit @ 2%)	25.12	26.76	25.94	5.59	6.63	6.11	
C <sub>9</sub> (Beejaumrit @ 3%)	24.86	26.63	25.74	5.41	6.53	5.97	
C <sub>10</sub> (Panchgavya @ 2%)	25.32	27.02	26.17	6.29	7.15	6.72	
C <sub>11</sub> (Panchgavya@3%)	24.22	26.69	25.46	5.68	6.57	6.13	
C <sub>12</sub> (Control)	20.40	23.87	22.14	4.89	5.34	5.12	
Mean	25.12	29.00		5.97	7.04		
SEm±		C 0.41 K 1.00 C×K 1.42		I	C 0.17 < 0.42 C×K 0.59		
CD (p=0.05)		C 0.63 K 1.57 C×K 2.21		(			

Chemicals	Number of leaves			Leaf area			
	K <sub>1</sub> (Intact mango stone)	K <sub>2</sub> (Extracted Kernel)	Mean	K <sub>1</sub> (Intact mango stone)	K <sub>2</sub> (Extracted kernel)	Mean	
C <sub>1</sub> (KNO <sub>3</sub> @ 0.5%)	8.40	11.65	10.03	78.83	70.86	74.85	
C <sub>2</sub> (KNO <sub>3</sub> @ 1%)	9.61	11.56	10.59	82.97	62.81	71.89	
C <sub>3</sub> (KNO <sub>3</sub> @ 1.5%)	7.62	10.60	9.11	69.39	84.55	76.97	
C <sub>4</sub> (BA @ 50 ppm)	7.66	12.59	10.13	67.56	70.32	68.94	
C <sub>5</sub> (BA @ 100 ppm)	7.06	13.00	10.03	63.97	78.97	71.47	
C <sub>6</sub> (GA <sub>3</sub> @ 100 ppm)	11.44	13.75	12.60	77.02	91.86	84.44	
C <sub>7</sub> (GA <sub>3</sub> @ 200 ppm)	10.08	13.99	12.03	70.85	100.61	85.73	
C <sub>8</sub> (Beejaumrit @ 2%)	8.04	12.00	10.02	58.05	70.89	64.47	
C <sub>9</sub> (Beejaumrit @ 3%)	9.15	11.01	10.08	65.11	68.26	66.69	
C <sub>10</sub> (Panchgavya @ 2%)	9.57	10.84	10.21	57.77	87.86	72.81	
C <sub>11</sub> (Panchgavya@3%)	8.47	10.08	9.28	59.07	89.93	74.50	
C <sub>12</sub> (Control)	8.10	6.86	7.48	51.04	53.09	52.06	
Mean	8.77	11.49		66.80	77.29		
SEm±		C 0.35 K 0.86 C×K 1.73		ŀ	1.42 3.48 C×K 4.93		
CD ( <i>p</i> =0.05)	1	C 0.72 K 1.76 C×K 2.48		C k	2.55		

Table 3: Effect of pre-soa	aking treatments of	on growth and su	rvival of see	dlings			
Chemicals	Seedling vigour index			Survival percentage			
	K <sub>1</sub> (Intact mango stone)	K <sub>2</sub> (Extracted Kernel)	Mean	K <sub>1</sub> (Intact mango stone)	K <sub>2</sub> (Extracted kernel)	Mean	
C <sub>1</sub> (KNO <sub>3</sub> @ 0.5%)	1610.8	2605.2	2108.1	65.69 (58.27)	69.31 (60.19)	67.50 (59.23)	
C <sub>2</sub> (KNO <sub>3</sub> @ 1%)	1583.7	2063.7	1823.7	68.59 (55.96)	75.21 (59.14)	71.90 (57.55)	
C <sub>3</sub> (KNO <sub>3</sub> @ 1.5%)	1467.4	2167.5	1817.5	72.19 (54.22)	72.69 (56.42)	72.44 (55.32)	
C <sub>4</sub> (BA @ 50 ppm)	1445.6	2329.2	1887.4	67.34 (55.24)	68.27 (55.79)	67.80 (55.51)	
C <sub>5</sub> (BA @ 100 ppm)	1432.7	2064.6	1748.6	60.55 (51.11)	77.31 (61.59)	68.93 (56.35)	
C <sub>6</sub> (GA <sub>3</sub> @ 100 ppm)	1925.2	3349.9	2637.4	78.35 (62.27)	82.53 (65.43)	80.44 (63.85)	
C <sub>7</sub> (GA <sub>3</sub> @ 200 ppm)	1759.8	2441.7	2100.8	70.65 (57.25)	80.56 (63.89)	75.60 (60.57)	
C <sub>8</sub> (Beejaumrit @ 2%)	1327.8	1651.9	1489.9	67.52 (55.31)	69.04 (56.19)	68.28 (55.75)	
C <sub>9</sub> (Beejaumrit @ 3%)	1364.6	1792.5	1578.6	63.04 (52.59)	72.12 (58.15)	67.58 (55.37)	
C <sub>10</sub> (Panchgavya @ 2%)	1447.1	2045.2	1746.2	62.96 (52.52)	70.59 (57.19)	66.77 (54.85)	
C <sub>11</sub> (Panchgavya@3%)	1443.6	1909.1	1676.3	69.69 (56.59)	69.30 (56.49)	69.50 (56.54)	
C <sub>12</sub> (Control)	974.4	1221.3	1097.8	47.98 (43.82)	56.38 (48.65)	52.18 (46.24)	
Mean	1481.9	2136.8		66.21 (54.60)	71.68 (58.26)		
SEm±	C	44.88; K 109.93 C×K 155.47			C 0.86; K 2.10 C×K 2.97		
CD ( <i>p</i> =0.05)		C 81.7; K 200 C×K 282.9			C 2.05; K 5.01 C×K NS		

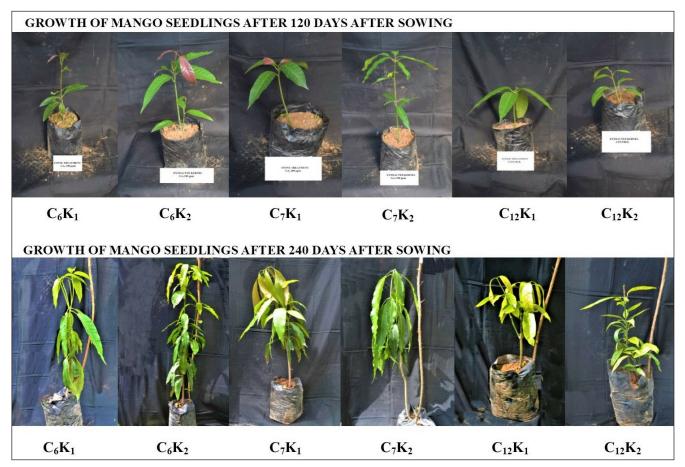


Figure 1: (Please mention figure title in correction table)

Maximum leaf area per leaf was recorded in seedlings raised form extracted kernels presoaked with GA<sub>3</sub> @ 200 ppm and these seed may have obtained maximum leaf area due stimulation of the cells on the use of GA<sub>3</sub>. As GA<sub>3</sub> at lower concentration was found effective in inducing division in cell causing the increase in the leaf area. The above results were found in close conformity with the reports provided by Kumar et al. (2008), Shaban (2010), Samir et al. (2016) in khrini, Vasantha et al. (2014) in tamarind.

Survival rate of seedling when calculated after 240 DAS was found maximum in extracted kernels treated with  ${\rm GA_3} \ @ \ 100$  ppm which can be attributed to the early germination of these seedlings as the time of sowing was august which was being followed by the harsh winter coming in October till February. The seedlings which have provided quick initiation were found having 80.44% survival in seedlings because the seedling had got enough time for establishing a better root network so as to tackle the harsh winter conditions. The results were found in confirmation with reports by Parmar et al. (2018) in jack fruit.

# 3.3. Vigour parameters

Vigor index is the distinct function of germination and growth, the maximum vigour index was recorded in seedlings which were raised from Extracted kernels treated with  ${\rm GA_3}$  @100 ppm however, this maximum vigour index can be justified as

the seedlings treated with GA<sub>3</sub> resulted production of more number of leaves per plant with maximum leaf area which led to production of high photosynthetic assimilate and providing best seedling height in turn producing the best seedling vigor index. The similar results were also reported by Padma and Reddy (1997), Aatla and Srihari (2013), Pampanna and Sulkieri (2001) in Sapota and Rajamanickam and Balakrishnan (2002) in anola.

### 4. Conclusion

Soaking of seeds in  $GA_3$  @ 100 ppm produced maximum (33.20 cm) plant height at 240 days. However, treatment of seeds with KNO $_3$  at lower doses was found promising in producing the highest proportion of graftable rootstocks and thus seedlings raised from extracted kernels pre-soaked in KNO $_3$  @ 0.5% attained graftable diameter within 210 days of sowing.

# 5. References

Aatla, H.B., Srihari, D., 2013. Influence of pre-sowing treatments on germination, growth and vigor of mango cv. Alphonso. Asian Journal of Horticulture 8, 122–125.
Bagul, H.B., Parmar, A.B., Parmar, B.K., Pandey, A.K., Jnanendra, M., 2018. Effect of pre-sowing treatment and growth of papaya (*Carica papaya* L.) seedlings cv. Red

- lady. The Pharma Innovation Journal 7, 95–97.
- Berjak, P., Pammenter, N.W., 2004. Recalcitrant seeds. In: Handbook of Seed Physiology (Benech-Arnold, R.A. and Sanchez, R.A.). Food Products Press, New York, pp. 305-345.
- Bewley, J.D., Black, M., 2012. Physiology and biochemistry of seeds in relation to germination: volume 2: viability, dormancy, and environmental control. Springer Science & Business Media.
- Brijwal, M., Kumar, R., Mishra, S., 2013. Effect of pre-sowing treatments on seed germination of guava (Psidium quajava L.) under Tarai region of Uttarakhand. Progressive Horticulture 45, 63–68.
- Choudhari, B.K., Chakrawar, V.R., 1981. Note on the effect of some chemicals on the germination of Rangpur lime seed. Indian Journal of Agricultural Science 51, 201–203.
- Dohono, W., Walker, R., 1957. Effect of gibberellic acid on breaking of rest period in Elberta peach. Science 126, 1178-1179.
- Gomez, A., 1984. Statistical procedures for agricultural research. 2<sup>nd</sup> ed. John Wiley and Sons, New York, 690p.
- Gupta, R., Chakrabarty, S.K., 2013. Gibberellic acid in plant: still a mystery unresolved. Plant signaling & behavior 8, 25504.
- Harris, D., Joshi, A., Khan, P.A., Gothkar, P., Sodhi, P.S., 1999. On-farm seed priming in semi-arid agriculture: development and evaluation in maize, rice and chickpea in India using participatory methods. Experimental Agriculture 35, 15–29.
- Kaur, S., Gupta, K., Huber, C., 1998. Gibberelin reverses the effect of salt stress in chickpea (Cicer arietinum L.) seedlings by enhancing amylase activity and mobilization of starch in cotyledons. Plant Growth Regulation 26, 85-90.
- Kozlowski, T.T., Pallardy, S.G., 1997. Growth control in woody plants. Elsevier. https://www.elsevier.com/books/ growth-control-in-woody-plants/roy/978-0-12-424210-4. ISBN:9780124242104
- Kumar, Y.H., 2007. Effect of pre-soaking treatments on germination, growth, vigour index and vigour of rootstocks in mango. Journal of Asian Horticulture 3, 157-161.
- Kumar, Y., Swamy, G.S.K., Kanmadi, V.C., Sowmaya, N., 2008. Effect of organics and chemicals on germination, growth and graft-take in mango. Asian Journal of Horticulture 3, 336-339.
- Loach, K., 1970. Shade tolerance in tree seedlings: II. Growth analysis of plants raised under artificial shade. New Phytologist 69, 273-286.
- Mandal, J., Mandal, B.K., Singh, R.R., Jaiswal, U.S., 2012. Effect of grafting height and cultivars on the performance of soft wood grafting in mango. Asian Journal of Horticulture 7, 171–174.
- Mauseth, J.D., 1976. Cytokinin-and gibberellic acid-

- induced effects on the structure and metabolism of shoot apical meristems in Opuntia polyacantha (Cactaceae). American Journal of Botany 63, 1295-1301.
- Mwaurah, P.W., Kumar, S., Kumar, N., Panghal, A., Attkan, A.K., Singh, V.K., Garg, M.K., 2020. Physicochemical characteristics, bioactive compounds and industrial applications of mango kernel and its products: A review. Comprehensive Reviews in Food Science and Food Safety 19, 2421–2446.
- Padma, M., Reddy, M., 1997. Effect of cracking and seed coat removal on seed germination of mango (Mangifera indica L.). Journal of Research-Acharya NG Ranga Agicultural University (India) 25, 18-21.
- Paleg, G., 1960. Physiological effects of Gibberellic acid II. On starch hydrolyzing enzymes of barley endosperm. Plant Physiology 35, 902.
- Pampanna, Y., Sulkieri, S., 2001. Effect of growth regulators on seed germination and seedling growth of sapota. Karnataka Journal of Agriculture Science 14, 1030–1036.
- Parmar, R., Parmar, P., Pandey, K., 2018. Effect of organic compounds on seed germination and seedling growth of jackfruit (Artocarpus heterophyllus Lam.) seed. The pharma innovation journal 7, 702-704.
- Patel, D., Gaikwad, S., Patel, D., 2016a. Effect of seed priming treatments on germination and seedling vigour of custard apple (Annona squamosa). Current Horticulture 4, 21–24.
- Patel, J., Ahlawat, R., Singh, A., Momin, S.K., Chaudhri, G., 2016b. Effect of pre-sowing treatments on stone germination and shoot growth of mango (Mangifera indica L.) seedlings. International journal of Agriculture Science 8, 2437-2440.
- Paulmurugan, S., 2021. "A Study on Trend and Growth of Mango in Tamil Nadu". Psychology and Education Journal 8, 6593–6600.
- Rajamanickam, C., Balakrishnan, K., 2002. Influence of seed treatments on seedling vigour in amla (Emblica officinalis G.). South Indian Horticulture 52, 324.
- Rajjou, L., Duval, M., Gallardo, K., Catusse, J., Bally, J., Job, C., Job, D., 2012. Seed germination and vigor. Annual Review of Plant Biology 63, 507–533.
- Ross, M.A., Harper, J.L., 1972. Occupation of biological space during seedling establishment. The Journal of Ecology 60, 77-88.
- Salisbury, B., Ross, C.W., 1992. Plant physiology. Belmont, California, Wads Worth Pub. Co, California. 154p.
- Samir, M., Rai, R., Parasad, B., 2016. Effect of organic manures on seed germination and seedling growth of Khirni. Indian Forester 142, 666–669.
- Sanchez, A., Soriano, A., Slabnik, S., 1967. The interaction of the seed coat and gibberellic acid in the germination of (Datura ferox L.). Canadian Journal of Botany 45, 371-376.
- Schnell, R., Brown, J.S., Olano, C., Meerow, A., Campbell,

- R., Kuhn, D., 2006. Mango genetic diversity analysis and pedigree inferences for Florida cultivars using microsatellite markers. HortScience 41, 993D-993.
- Shaban, A., 2010. Improving seed germination and seedling growth of some mango rootstocks. American-Eurasian Journal of Agricultural and Environmental Science 7, 535-541.
- Sharaf, M.M., Atawia, A.R., Bakry, K.A., EL-Rouby, M.Z., 2016. Effect of pre-sowing seeds soak in different GA<sub>3</sub> and ZnSO, solutions on germination and growth of Cleopatra mandarin and Rangpur lime rootstocks. Middle East Journal of Agricultural Research 5, 233-238.
- Singh, N.K., Mahato, A.K., Jayaswal, P.K., Singh, A., Singh, S., Singh, N. and Sharma, T.R. 2016. Origin, diversity and

- genome sequence of mango (Mangifera indica L.). Indian Journal of History of Science 51, 355–368
- Tharanathan, R.N., Yashoda, H.M., Prabha, T.N., 2006. Mango (Mangifera indica L.), "The king of fruits"—An overview. Food Reviews International 22, 95–123.
- Umarani, R., Aadhavan, E.K., Faisal, M.M., 2015. Understanding poor storage potential of recalcitrant seeds. Current Science 108, 2023–2034.
- Vasantha, P.T., Vijendrakumar, R.C., Guruprasad, T.R., Mahadevamma, M., Santhosh, K.V., 2014. Studies on effect of growth regulators and biofertilizers on seed germination and seedling growth of tamarind (Tamarindus indica L.). Plant Archives 14, 155-160.