



Assessment of Heterosis for Root Yield and Attributing Traits in Ashwagandha [*Withania somnifera* (L.) Dunal]

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ABSTRACT

A study was conducted to estimate the extent of heterosis for root yield and attributing traits in ashwagandha at the Botanical Garden, Department of Genetics and Plant Breeding, C. P. C. A, S. D. Agricultural University, Sardarkrushinagar, Gujarat, India. Thirty six hybrids were generated from the diallel mating design of 9 diverse parents during October 2018 – March 2019 (*rabi* season). Forty five entries were evaluated in RBD with 3 replications during October 2019–March 2020 (*rabi* season) for the yield and attributing traits. The root traits are given more emphasis because of its economic value. All the forty-five genotypes exhibited moderate to good mean performance along with an adequate amount of variability among and between the parents and hybrids. Most of the hybrids showed highly significant better parent and standard heterosis in desirable direction for the traits considered. The hybrids like SKA 10×JA 20, JA 134×AWS 1, SKA 24×SKA 26, SKA 24×AWS 1, SKA 11×AWS 1, SKA 11×SKA 26 and JA 20×AWS 1 were with a markedly significant heterotic response for fresh root yield trait. The results also suggest that the cross JA 134×AWS 1 is appropriate to exploit heterosis in root length, fresh root yield and dry root yield. Further evaluation and generation advancement of these successful crosses can derive transgressive segregants and hybrids with high yield potential for commercialization.

KEYWORDS: Ashwagandha, diallel analysis, heterosis, hybrids, standard heterosis

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Data Availability Statement: Legal restrictions are imposed on the public sharing of raw data. However, authors have full right to transfer or share the data in raw form upon request subject to either meeting the conditions of the original consents and the original research study. Further, access of data needs to meet whether the user complies with the ethical and legal obligations as data controllers to allow for secondary use of the data outside of the original study.

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1. INTRODUCTION

Ashwagandha [*Withania somnifera* (L.) Dunal] is a Solanaceous medicinal plant used in diverse medicinal systems like Ayurveda, Siddha and Unani. This quality herb possesses therapeutic value against a number of ailments such as arthritis, asthma, mental diseases, inflammation, rheumatism, infections, fever, anxiety, tuberculosis, male sexual disorders and cancer (Mirjalini et al., 2009; Hahm and Singh, 2013; Gosh and Halder, 2015). Different parts of the ashwagandha plant contain a number of chemical compounds among which the alkaloids or more specifically the withanolides hold immense importance from pharmacological point of view. The withanolides present in ashwagandha plants are species specific. *W. somnifera* and *W. ashwagandha* contains withaferin A as major compound, in contrast coagulin L is present in *W. coagulans*. Beside these this plant contains amino acids including aspartic acid, proline, tyrosine, glutamic acid, cystine, tryptophan, alanine, and elevated amount of iron. Root, leaves, fruits and seeds are the commercial parts of ashwagandha that possess medicinal properties but roots are the major commercial part which contains 0.4–1.2% alkaloids, 40–65% starch, 40–65% fibres and a minor quantity of oil (Joshi et al., 2014). Apart from the medicinal properties ashwagandha root powder is directly used as an ingredient in different recipes like fruit-juice beverages, bakery items, savoury, and snack items.

Two out of twenty-six species of *Withania*, namely *W. somnifera* and the wild *W. coagulans* are found in India (Kaul et al., 2005). A third species namely *W. ashwagandha* is also delineated from *W. somnifera* based on morphological, chemical and molecular studies (Kaul, 1957; Mir et al., 2010). The name Ashwagandha is originated from the Sanskrit language and is a combination of “Ashwa” means horse and “gandha” means smell (Murthy et al., 2010). It is because of the smell of this plant’s root is comparable to horse sweat. The plant is native to dry regions of India, Australia, East Asia, and Africa. It grows well in a dry climate and low winter temperatures because of hardiness and drought tolerance. It is grown as a late rainy season crop as the late winter rains facilitate proper root development (Joshi et al., 2014). Major cultivated areas of India include Madhya Pradesh, Rajasthan, Gujarat, Punjab and Uttar Pradesh. Ashwagandha cultivation is a better choice for lands which is not suitable for food crops (Kothari et al., 2003). The area under the Ashwagandha crop is 1461 ha according to the 2015–16 cultivation status of Anonymous (2015–16). The annual root production in India is estimated to be about 2000 tons (Kumar et al., 2010).

The breeding works for the improvement of the ashwagandha crop are limited. Even though, some varieties were released by CSIR–CIMAP, Lucknow including Poshita, Rakshita

and Pratap. Jawahar 20 is a high yielding variety released from Madhya Pradesh (Khare et al., 2020). WSR is another popular variety released by CSIR–Regional Research Laboratory, Jammu. Variety GAA 1 was released in the year 2015 for Gujarat state by AAU, Anand. The plant flowers and sets fruits during November to February. The floral architecture of both cultivated and wild populations of Ashwagandha creates conditions favouring selfing (Kaul et al., 2005). Natural variability of self-pollinated populations tends to exhaust due to selection and hybridization become necessary for further genetic improvement. The increased demand for ashwagandha in the drug industry (necessitating the development of high yielding hybrids and genetic analysis of this crop. Taking these facts into account, an experiment was conducted to record the extent of heterosis for root yield and its attributing traits.

2. MATERIAL AND METHODS

2.1. Experimental material

The experimental material consisted of nine parental lines (SKA 27, SKA 11, SKA 6, SKA 24, SKA 10, SKA 26, JA 20, JA 134 and AWS 1) and the hybrids generated by half-diallel mating design. Details of parental lines are arranged in Table 1.

Table 1: Details of parental lines

Sl. No.	Name of parental line	Pedigree	Source
1.	SKA 27	IPS from MPAS 7	S. K. Nagar
2.	SKA 11	IPS from MWS 101	S. K. Nagar
3.	SKA 6	IPS from MWS 322	S. K. Nagar
4.	SKA 24	IPS from MPAS 3	S. K. Nagar
5.	SKA 10	IPS from MWS 309	S. K. Nagar
6.	SKA 26	IPS from MPAS 5	S. K. Nagar
7.	JA 20	–	Mandsaur, Madhya Pradesh
8.	JA 134	Pedigree selection from JA 20 and wild types	Mandsaur, Madhya Pradesh
9.	AWS 1 (Check)	–	AAU, Gujarat

2.2. Crossing programme

Crosses were attempted during *rabi* season (October, 2018–March, 2019) at the Botanical Garden, Department of Genetics and Plant Breeding, C. P. C. A, S. D. Agricultural University, Sardarkrushinagar, Gujarat (latitudes of 24°.31' N, longitude of 72°.32' E and altitude of 154.52 meters above MSL).



The study on pollen fertility and stigma receptivity in closed flowers of *W. somnifera* (L.) shows the flower is kind of chasmogamous (Lattoo et al., 2007) and the time duration between 7.00 to 11.00 hours are most suitable for 95–98% success in hybridization. Mature flower buds of female parents about to open next day morning were selected for hand emasculation. It was carried out between 3.00 to 5.00 p.m by removing the calyx and anthers by using forceps without damaging pistils. The emasculated buds were bagged to avoid open pollination. Pollination was done next day morning between 8.00 to 11.00 am by rubbing the dehisced flower on the stigma of the emasculated flower. Pollinated flowers were bagged immediately with proper labelling. Tagging was done after the berry set and mature berries of crosses were collected separately. Seeds were stored in proper conditions after drying and threshing.

2.3. Evaluation and recording of observations

The 36 crosses generated along with their 9 parental lines were evaluated in Randomized Block Design with three replications during *rabi* 2019–2020 at Agronomy Instructional Farm, S. D. Agricultural University, Sardarkrushinagar. Each replication contained one row of each 45 entries. A row length of 4 m and a spacing of 30×10 cm was maintained. Observations were recorded for ten yield and attributing traits such as days to flowering, plant height (cm), number of primary branches, number of secondary branches, number of berries plant⁻¹, number of seeds berry⁻¹, root length (cm), root girth (cm), fresh root yield (g) and dry root yield (g).

2.4. Statistical analysis

The replication wise mean values of all these treatments were statistically analysed as per procedures suggested by Panse and Sukathme (1985). The magnitude of heterosis over better parent (Heterobeltiosis) and standard check (Standard Heterosis) were estimated as per Fonseca and Patterson (1968) and Meredith and Bridge (1972) respectively. The variety AWS–1 was considered as a check for the estimation of standard heterosis.

3. RESULTS AND DISCUSSION

Identification of crosses with significant heterotic responses for root yield and attributing traits has immense importance in the genetic improvement of ashwagandha. This experiment targeted studying the extent of heterobeltiosis and standard or economic heterosis which is relevant to practical plant breeding.

The analysis of variance of the present investigation showed significance among the mean sum of squares due to genotypes for all traits under study (Table 2). Kumar et al. (2007), Mir et al. (2014), Srivastava et al. (2018) and Dhaka et al. (2021) reported the presence of a considerable amount of variation in various traits of ashwagandha. The mean sum of squares due to parent vs hybrids of the present study was highly significant for all traits except days to flowering and dry root yield (g) implied the marked difference in the performances of cross combinations among them and compared to the parental lines. This available variation in the population is suitable to perform selection and to create

Table 2: Analysis of variance for root yield and attributing traits in Ashwagandha

Sources of variation	d. f.	Days to flowering	Plant height (cm)	No. of primary branches	No. of secondary branches	No. of berries plant ⁻¹	No. of seeds berry ⁻¹	Root length (cm)	Root girth (cm)	Fresh root yield (g)	Dry root yield (g)
Replications	2	36.07	16.07*	0.04	2.37*	110.56	20.16	32.85*	0.59	1.89	0.21
Genotypes	44	174.30**	249.15**	3.69**	7.11**	27008.35**	108.35**	100.95**	3.70**	41.00**	2.02**
Parents	8	107.04**	154.26**	1.06**	6.39**	4847.18**	111.33**	111.86**	2.54**	48.69**	3.33**
Hybrids	35	194.59**	228.10**	4.00**	6.43**	27733.66**	92.11**	99.31**	3.84**	39.73**	1.77**
Parent vs hybrids	1	2.14	1744.78**	14.07**	36.84**	178911.83**	625.87**	71.25**	8.10**	24.01**	0.12
Error	88	13.82	4.93	0.18	0.30	125.23	12.65**	4.55	0.30	1.99	0.14

* $p \leq 0.05$, ** $p \leq 0.01$

heterotic combinations using diverse parents. Reliable results showing the existence of variance in parents and heterotic combinations in ashwagandha for the one or more traits of the present study are evident from Dhuri (2016).

High mean performance is always a consideration in the selection of parental lines and the creation of

heterotic groups. All the nine parents and 36 hybrids displayed moderate to good performance for traits under consideration. Regarding the root traits, the parents SKA 27, SKA 6 and SKA 10 were identified to be with the best performance. The majority of the hybrids exhibited superior mean performance compared to parents (mean performance



of the entries is attached as supplementary information). The majority of the crosses showed greater magnitudes of heterosis in the desired direction over better parent and the check (AWS 1) for root yield and its attributing traits. The details including a range of heterosis estimates and the number of crosses with significance in a positive and negative direction are arranged in Table 3. The estimates of better parent and standard heterosis over the check AWS 1 for all the 36 crosses generated are given in the supplementary information.

The hybrids with negative significant heterosis are agronomically desirable in ashwagandha for days to flowering and plant height. Out of ten negatively significant hybrids, SKA 11×SKA 26 (−14.73%), SKA 24×AWS 1 (−14.54%), SKA 11×JA 20 (−13.69%), JA 20×AWS 1 (−10.79%) and SKA 10×JA 134 (−9.28%) showed highly significant negative heterobeltiosis. The hybrids showed highly negative significance over the check AWS 1 includes SKA 10×SKA 26 (−17.46%), SKA 26×JA 134 (−17.05%), SKA 27×SKA 6 (−15.38%) and SKA 11×JA 20 (−13.30%).

Table 3: A summary of heterosis analysis for traits in Ashwagandha

Traits	Better parent heterosis (%)			Standard heterosis (%)		
	Range	No. of positive significant crosses	No. of negative significant crosses	Range	No. of positive significant crosses	No. of negative significant crosses
DF	−14.73 to 28.37	20	10	−17.45 to 14.21	17	14
PH	−31.58 to 88.21	33	1	−48.60 to 15.41	6	22
PB	−42.11 to 143.16	4	–	−44.27 to 114.56	2	–
SB	−37.87 to 84.08	5	1	−42.45 to 77.33	6	–
BPP	44.70 to 253.90	27	8	−64.68 to 127.89	17	19
SPB	−26.90 to 36.42	17	8	−11.96 to 52.29	29	3
RL	−52.61 to 99.21	12	18	−18.51 to 92.70	20	3
RG	−89.77 to 73.28	2	2	−25.99 to 73.29	7	–
FRY	−78.56 to 146.50	9	13	−28.37 to 156.61	21	–
DRY	−86.15 to 120.66	1	3	3.52 to 135.24	3	–

DF: Days to flowering; PH: Plant height; PB: Number of primary branches; SB: Number of secondary branches; RL: Root length; RG: Root girth; BPP: Number of berries plant^{−1}, SPP: Number of seeds^{−1}; DRY: Dry root yield

For the trait plant height, only one cross exhibited negative significance over the better parent namely SKA 24×JA 134 (−31.58%). Standard heterosis over the check AWS 1 found to be manifested in 22 crosses which includes SKA 24×JA 134 (−48.61%), SKA 10×SKA 26 (−38.55%), SKA 6×SKA 24 (−34.57%), SKA 11 × SKA 6 (−33.55%) etc. These results are in agreement with the findings of Arpan et al. (2014) and Sharma et al. (2016).

As the number of primary branches increases the number of berries as well as the seed yield increases. The top crosses that showed high positive significance for heterobeltiosis were SKA 24×SKA 26 (143.16 %), SKA 10×JA 20 (105.64%) and SKA 26×AWS 1 (14.12%). Highly significant positive heterotic crosses over the standard check include SKA 10×JA 20 (114.56%) and SKA 26×AWS 1 (92.12%). The number of secondary branches was found to be correlated with the root length, diameter and fresh weight as per the findings of Arpan et al. (2014). Hence the significant heterosis for the number of secondary branches can take

into account for the exploitation of hybrid vigour in the root traits as well. Four of the fourteen significant hybrids showed the heterobeltiosis in a positive direction in the case of the number of secondary branches. Crosses like SKA 2×AWS 1 (77.33%) and SKA 27×SKA 24 (67.25%) were with high magnitude and positive significance in the contest of economic heterosis.

For the number of berries plant^{−1} twenty-seven crosses were found to be significant over the better parent. The crosses SKA 27×SKA 26 (253.90%), SKA 27×SKA 11 (242.27%) and SKA 24×SKA 26 (214.76%) showed higher magnitudes of heterobeltiosis. Seventeen hybrids were positively significant over the check AWS 1. The cross SKA 27×SKA 24 (127.89%) exhibited the highest positive significance for standard heterosis followed by SKA 27×SKA 11 (114.98%), SKA 10×JA 134 (99.29%) and SKA 11×SKA 26 (84.11%). Twenty-five crosses out of the thirty-six crosses showed significance over the better parent for the trait number of seeds berry^{−1}. Among that, twenty-three crosses were



highly significant in a positive direction. A broad range of standard heterosis was observed for this trait and the crosses like SKA 27×JA 134 (52.29%), SKA 27×SKA 24 (52.05%), SKA 6×SKA 24 (49.18%), SKA 10×JA 134 (47.55%) and SKA 6×SKA 26 (45.62%) manifested significant heterosis in a positive direction.

Root traits are crucial to the ashwagandha crop. The results for root length traits indicated that 30 crosses manifested significant heterosis over their respective better parents which includes SKA 27×SKA 11 (99.21%), SKA 24×AWS 1 (92.69%), SKA 27×AWS 1 (62.32%), SKA 27×SKA 24 (57.27%), SKA 11×JA 20 (49.36%) and SKA 6×AWS 1 (28.31%). Twenty hybrids showed highly significant positive heterosis over the check. The crosses like SKA 24×AWS 1 (92.70%), SKA 27×SKA 11 (66.69%), SKA 11×SKA 6 (66.26%), SKA 27×AWS 1 (62.32%), SKA 27×SKA 24 (56.15%) and SKA 11×SKA 26 (54.11%) showed highly significant standard heterosis. The estimates for root girth implied that only two crosses viz., SKA 11×JA 20 (73.28%) followed by SKA 24×AWS 1 (56.86%) were with significant heterobeltiosis in a positive direction. Top hybrids with the highest positive standard heterosis was SKA 11×JA 20 (79.29%) followed by SKA 6×SKA 26 (67.9%), SKA 27×AWS 1 (65.94%) and SKA 10×JA 20 (61.28%).

A wide range of heterosis was exhibited by the hybrids generated for the most important traits like fresh root yield and dry root yield. Eight crosses displayed positive heterosis for fresh root yield over the better parent of which JA 134×AWS 1 (146.50^{***}) followed by SKA 24×SKA 26 (97.75^{***}) and SKA 24×AWS 1 (89.95^{***}) exhibited the highest magnitude. Significant positive heterosis for fresh root yield was also observed for 15 crosses. Among that, the top three crosses were SKA 10×JA 20 (156.62^{***}), SKA 6×SKA 26 (153.19^{***}) and SKA 24×SKA 26 (90.46^{**}). The perusal estimates of heterobeltiosis for dry root yield indicated that only one cross viz., JA 134×AWS 1 (120.66^{***}) was highly significant in a positive direction over the better parent. The crosses JA 134×AWS 1 (135.25^{***}), SKA 27×SKA 10 (125.36^{*}) and SKA 10×SKA 26 (101.33^{*}) displayed greater magnitudes for standard heterosis.

The results derived from this study points towards the possibility of exploitation of heterosis in the ashwagandha germplasm for the improvement of root yield related traits which is in agreement with the finding of Patidar et al. (1990), Arpan et al. (2014) and Dhuri (2016). Accordign to previous reports the heterotic combinations for root yield is expected to show high total alkaloid content due to significant correlation (Kumar et al., 2011).

Hybrid vigour of the root traits is of great value since root is the major economic part of ashwagandha. Many of the significant heterotic crosses for fresh root yield revealed

in this study exhibited considerable extend of heterosis in desirable direction for other attributing traits as well. The cross combinations given in Table 4 are appropriate to utilize according to the traits in which heterosis is desired in ashwagandha.

Table 4: Superior heterotic combinations for fresh root yield

S l. No.	Crosses	Better parent heterosis (%)	Standard heterosis (%)	Other traits with heterosis in the desired direction
1.	SKA 10×JA 20	156.62 ^{**}	52.80 ^{**}	PH, PB, BPP, SPB, RL, RG
2.	JA 134×AWS 1	150.90 ^{**}	146.50 ^{**}	PH, SPB, RL, DRY
3.	SKA 24×SKA 26	90.46 ^{**}	97.75 ^{**}	BPP, SPB
4.	SKA 24×AWS 1	89.95 [*]	89.95 ^{**}	DF, BPP, SPB, RL, RG
5.	SKA 11×AWS 1	84.48 ^{**}	53.28 ^{**}	DF, PH, SB, BPP, RL, RG
6.	SKA 11×SKA 26	9381.9495 ^{***}	88.90 ^{**}	SB, BPP, SPB, RL
7.	JA 20×AWS 1	54.46 [*]	28.33 [*]	DF, SPB, RL

DF: Days to flowering; PH: Plant height; PB: Number of primary branches; SB: Number of secondary branches; RL: Root length; RG: Root girth; BPP: Number of berries plant⁻¹, SPP: Number of seeds⁻¹; DRY: Dry root yield

4. CONCLUSION

This study shows the possibility of heterosis breeding in ashwagandha. The crosses SKA 10×JA 20, JA 134×AWS 1, SKA 24×SKA 26, SKA 24×AWS 1, SKA 11×AWS 1, SKA 11×SKA 26 and JA 20×AWS 1 are useful to exploit heterosis for fresh root yield. Among these JA 134×AWS 1 could be a better choice for hybrid development due to the significant heterotic response in root length, fresh root yield and dry root yield.

5. FURTHER RESEARCH

These hybrids can be advanced over generations to derive transgressive segregants and information about genetic components of variance. Further field evaluations and molecular analyses are suggested for the release of potential hybrids in ashwagandha.



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