



Variability on Seed Yield and Related Agronomic Traits in Mesoamerican Common Bean (*Phaseolus vulgaris* L.) Genotypes under Contrasting Environments at South Eastern Ethiopia

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ABSTRACT

The experiment was conducted during July–November, 2022 at Melkassa and Kulumsa Agricultural Research Center main station under rain fed condition to assess the extent of genetic variability and to identify potential candidate genotypes. A total of 64 small seeded common bean genotypes were evaluated for 18 quantitative traits using 8*8 triple lattice design. The analysis of variance showed that there was highly significant ($p \leq 0.01$) difference among genotypes for all studied traits at both individual locations. The genotypes variation for seed yield was ranged from 1685.09 kg ha⁻¹ to 4499 kg ha⁻¹ at Melkassa and from 1369.76 kg ha⁻¹ to 4848.38 kg ha⁻¹ at Kulumsa. High PCV and GCV were recorded for total number of seeds per plant, number of fertile pods and total number of pods plant⁻¹ in both locations. Moderate to high value of GCV and broad sense heritability coupled with high GAM were obtained for seed per plant, dry pod yield per plant, fertile pods per plant, seed weight per plant, hundred seed weight and seed yield per hectare in both locations; indicating the importance of those traits in yield improvement of small seeded common bean. G-39, G-27, G-58, G-35, G-29, G-18, G-8, G-61, G-37 and G-33 showed yield advantage from 49% to 88.9% at Melkassa and G-33, G-8, G-4, G-12, G-58, G-11, G-45, G-13, G-36 and G-30 from 16% to 36% at kulumsa over the best standard check G-64. The existed variations implied the possibility for further improvement of seed yield and atributing traits through utlizing selected potential genotypes and targeted hybridization schemes in breeding programs.

KEYWORDS: Crop improvement, genetic advance, genotype, heritability, variability

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1. INTRODUCTION

Common bean (*Phaseolus vulgaris* L.) is one of the most important warm season food legume crop belongs to the fabaceae family (Anonymous, 2016). It is a global food legume crop grown on 34.8 mha of land with annual total production of over 27.5 mt (Anonymous, 2020). Common bean is the second most important food legumes in Africa next to faba bean as a source of dietary protein (Broughton et al., 2003). Currently in Ethiopia, common bean is the second major food legume crop after faba bean in terms of both area and total amount of production (Anonymous, 2019). The production and area allocation for common bean in Ethiopia is steadily increasing from time to time. In 2018/19 main cropping season more than 2.8 million smallholders are engaged in common bean production including red and white bean. It is grown on 288,637.23 ha of land with total production of 488,320.17 tons; which accounts for 17.81% of the total area covered by pulses and 16.21% of the total pulses production in the country in the main growing season (Anonymous, 2019).

It has great versatile purposes in the livelihood of the agricultural societies of the country. It serves as a source of food consumed as *Nifro* (boiled grain mixed with sorghum or maize), *Shirowat*, *soup*, *samosa* and the immature pods and seeds as a vegetable. The crop is also used as a rotational crop in cereal based cropping systems for sustainable production due to its pertinent atmospheric nitrogen fixing. In addition, common bean is important as a source of animal feed, cash to the farmers and foreign currency earning to the country.

Despite its huge importance in the country, the national average productivity of the crop (1.72 t ha^{-1}) (Anonymous, 2019) is low compared to the yield potential of the crop (3.5 t ha^{-1}) (Anonymous, 2017; Berhanu et al., 2018). These is primarily due to limited availability of source materials and improved varieties for stress tolerant such as biotic stresses like diseases, insect pests, weeds and abiotic factors such as soil acidity, drought and, instability of cultivars across different agro-ecology, poor adaptation and poor crop management (Ketema and Thangavel, 2016; Berhanu et al., 2018).

On the other hand, currently, in Ethiopia the demand for improved common bean varieties especially for small red and white colored beans among producers are increasing

as a result of the increasing demand of consumers and marketing industry (Frehiwot, 2010; Ephrem, 2016) even though the producers have varied preferences in different geographical regions of Ethiopia (Yonas, 2017). Hence, further development of desirable genotypes are essential through plant breeding program for selection of superior genotypes. These depend upon the extent of genetic variability and genetic advance in the base population with respect to desired traits.

There are many research studies have been conducted in Ethiopia on genetic variability in common bean (Ejigu et al., 2018, Aziza, 2019). However, limited research efforts have been done on genetic variability for genotypes of different seed size groups separately; while there are still numerous introduced and locally crossed common genotypes in which their genetic variability does not properly and systematically studied. In addition, information on genetic variability for small seeded genotypes considered in this study has not been generated so far. Therefore, the present study was conducted under contrasting environments (low and mid altitude areas) to estimate the extent of variability, heritability, and the expected genetic advance of important morpho-agronomic traits and to identify potential candidate genotypes to be used for future breeding activity.

2. MATERIALS AND METHODS

2.1. Experimental site

The experiment was conducted at two locations of South Eastern Ethiopia namely Melkassa and Kulumsa Agricultural Research Center main station during the main cropping season in July–November 2022 under rain fed condition. The description of the test environments are shown in Table 1.

2.2. Experimental materials and design

Sixty-four small seeded common bean genotypes, including sixty-two breeding lines and two recently national released varieties were used for the study. List of common bean genotypes, code and origin are given in Table 2. The experiment was carried out using 8×8 triple lattice design; each replication containing eight incomplete blocks and each incomplete block containing eight genotypes. Each plot had four rows of 4 m length, with spacing of 40 cm between rows and 10 cm between plants. Each genotype was planted in a plot size of 6.4 m^2 with 3.2 m^2 for net harvesting.

Table 1: Description of the test environments

| Testing location | Altitude (m.a.s.l) | Latitude | Longitude | Annual Rainfall (mm) | Min.annual Temp. (°C) | Max.annual Temp. (°C) |
|------------------|--------------------|-------------|--------------|----------------------|-----------------------|-----------------------|
| Melkassa | 1550 | 8°25' N | 39° 20' E | 763 | 16 | 28.8 |
| Kulumsa | 2200 | 8° 01' 10"N | 39° 09' 13"E | 850 | 7.9 | 23.1 |



Table 2: List of genotypes used in the experiment

| Code | Genotype | Source of origin | status | Code | Genotype | Source of origin | status |
|------|---------------|------------------|---------------|------|---------------|------------------|--------------------|
| G-1 | SSIN1148 | CIAT | breeding line | G-33 | NUA355 | CIAT | breeding line |
| G-2 | NUA648 | CIAT | breeding line | G-34 | SCAM15-21-124 | CIAT | breeding line |
| G-3 | SMR106 | CIAT | breeding line | G-35 | GENO363 | MARC | breeding line |
| G-4 | GENO110 | MARC | breeding line | G-36 | SMR126 | CIAT | breeding line |
| G-5 | SEC22 | CIAT | breeding line | G-37 | SSIN1347 | CIAT | breeding line |
| G-6 | SCAM15-21-348 | CIAT | breeding line | G-38 | GENO206 | MARC | breeding line |
| G-7 | GENO161 | MARC | breeding line | G-39 | SSIN1020 | CIAT | breeding line |
| G-8 | CB170064-5 | MARC | breeding line | G-40 | SCAM15-21-357 | CIAT | breeding line |
| G-9 | GENO188 | MARC | breeding line | G-41 | SMR44 | CIAT | breeding line |
| G-10 | CB170072-13 | MARC | breeding line | G-42 | GENO34 | MARC | breeding line |
| G-11 | SSIN939 | CIAT | breeding line | G-43 | CB170065-21-1 | MARC | breeding line |
| G-12 | SMR54 | CIAT | breeding line | G-44 | GENO 354 | MARC | breeding line |
| G-13 | GENO285 | MARC | breeding line | G-45 | SER347 | CIAT | breeding line |
| G-14 | GENO147 | MARC | breeding line | G-46 | SMR48 | CIAT | breeding line |
| G-15 | CB170058-11 | MARC | breeding line | G-47 | SCAM15-11-154 | CIAT | breeding line |
| G-16 | SCAM15-21-125 | CIAT | breeding line | G-48 | GENO126 | MARC | breeding line |
| G-17 | GENO158 | MARC | breeding line | G-49 | SSIN1358 | CIAT | breeding line |
| G-18 | CB170065-22-2 | MARC | breeding line | G-50 | SMR95 | CIAT | breeding line |
| G-19 | GENO245 | MARC | breeding line | G-51 | GENO418 | MARC | breeding line |
| G-20 | SMR123 | CIAT | breeding line | G-52 | SCAM15-21-381 | CIAT | breeding line |
| G-21 | SSIN1309 | CIAT | breeding line | G-53 | SMR103 | CIAT | breeding line |
| G-22 | GENO214 | MARC | breeding line | G-54 | SMR83 | CIAT | breeding line |
| G-23 | SSIN885 | CIAT | breeding line | G-55 | SSIN1313 | CIAT | breeding line |
| G-24 | GENO45 | MARC | breeding line | G-56 | GENO276 | MARC | breeding line |
| G-25 | CB170044-13-3 | MARC | breeding line | G-57 | GENO122 | MARC | breeding line |
| G-26 | GENO341 | MARC | breeding line | G-58 | CB170065-51-1 | MARC | breeding line |
| G-27 | SCAM15-21-430 | CIAT | breeding line | G-59 | SCAM15-21-227 | CIAT | breeding line |
| G-28 | SMR53 | CIAT | breeding line | G-60 | GENO331 | MARC | breeding line |
| G-29 | GENO263 | MARC | breeding line | G-61 | CB170044-91-1 | MARC | breeding line |
| G-30 | SMR46 | CIAT | breeding line | G-62 | GENO66 | MARC | breeding line |
| G-31 | SSIN956 | CIAT | breeding line | G-63 | RAZ42 | MARC | Commercial variety |
| G-32 | GENO186 | MARC | breeding line | G-64 | SCR15 | MARC | Commercial variety |

Where: MARC: Melkassa Agriculture Research Center; CIAT: International Center for Tropical Agriculture

2.3. Data collection

Data were collected on single plant and plot bases. On a plant basis, data were collected from ten randomly selected plants from each genotype in each replication, namely, plant height (PH) (cm), pod length (PL) (cm), pod diameter (PD) (mm), total number of pods per plant (TNPPP) (number), number of fertile pods plant⁻¹ (NFPPP) (number), dry

pod yield plant⁻¹ (DPYPP) (g plant⁻¹), single pod dry weight (SPDW) (g pod⁻¹), number of seeds pod⁻¹ (NSPP) (number), total number of seeds plant⁻¹ (TNSPP) (number), and seed weight plant⁻¹ (SWPP) (g plant⁻¹). While the data on plot basis were collected from the two central rows include stand count at emergence (SCE), days to 50% flowering (DF), days to 90% maturity (DM), stand count



at harvest (SCH), above ground biomass yield (BMYPH) (kg ha⁻¹), hundred seed weight (HSW) (gram), seed yield (SYPH) (kg ha⁻¹) and harvest index (HI) (%).

2.4. Data analysis

the analysis of variance was carried out using the procedure of triple lattice design for all traits to assess the significance of the difference among the genotypes by using lmer function of stats package in R software version 4.1. The genotypic variance (σ^2_g) and the environmental variance (σ^2_e) were obtained directly from variance component table generated by the software using lmer function of the lme4 mixed model package in R software version 4.1.2 by considering the genotype in the linear mixed model as random and replication and block as fixed variable using Residual (restricted) maximum likelihood (REML) variance component estimation method. Other genetic parameters were calculated by Microsoft Excel using suggested equations by (Falconer and Mackay, 1996; Johnson et al., 1955).

Genotypic variance (σ^2_g) = (MSg - MSe) / r

Environmental variance (σ^2_e) = MSe

Phenotypic variance (σ^2_p) = $\sigma^2_g + \sigma^2_e$

Where: MSg = Mean square of genotypes

Mse = Mean square of error

r = Number of replication

Phenotypic and genotypic coefficients of variations of each trait were expressed as percentage of the corresponding phenotypic and genotypic standard deviations as described by Johnson et al. (1955) and expressed as follows:

PCV = $\sqrt{(\sigma^2_p / \bar{x})} \times 100$

GCV = $\sqrt{(\sigma^2_g / \bar{x})} \times 100$

Where: PCV = Phenotypic coefficient of variation, GCV = Genotypic coefficient of variation, σ^2_p = Phenotypic variance, σ^2_g = Genotypic variance, \bar{x} = mean value of the trait

According to Deshmukh et al. (1986) the PCV and GCV estimates classified as low, <10%, Moderate, 10–20%, High, >20%

Broad sense heritability values for all parameters (H^2_B) were estimated based on the formula given by (Falconer and Mackay, 1996).

$H^2_B = \sigma^2_g / (\sigma^2_p) \times 100$

Where: H^2_B = Heritability in broad sense, σ^2_g = Genotypic Variance, σ^2_p = Phenotypic variance

Heritability estimates in broad sense was categorized as high (>60%), medium (30–60%) and low (<30%) (Dabholkar, 1992).

Genetic advance (GA) at 5% selection intensity was estimated as per formula given by (Johnson et al., 1955)

$GA = K \times \sqrt{(\sigma^2_p)} \times H^2_B$

Where: GA = Genetic advance, K = Selection differential at 5% selection intensity which accounts to a constant value 2.063, σ^2_p = Phenotypic variance, H^2_B = Broad sense heritability

The genetic advance as percent of mean (GAM) was calculated using the following formula and was expressed in percentage (Johnson et al., 1955).

$GAM = (GA / \bar{x}) \times 100$

Where: GAM = Genetic advance as percent of mean, GA = Genetic advance at 5% selection intensity, \bar{x} = population Mean

According to Johnson et al. (1955), the GAM was classified as low if <10%, Moderate, 10–20% and High, >20%

3. RESULTS AND DISCUSSION

The analysis of variance showed the presence of highly significant ($p \leq 0.01$) difference among 64 common bean genotypes for all studied traits at both locations (Table 3 and 4). The presence of highly significant difference was an indication of the existence of considerable genetic variability in experimental materials for the studied traits, justified carrying out further genetic analysis. This will provide an opportunity for a breeder to select superior and desired genotypes for their better seed yield and other yield related traits for further improvement through exploiting the observed variations. Different researchers reported significant differences for one or more of the studied traits (Aziza, 2019; Abnet, 2020; Gebeyaw et al., 2021).

The present study indicated that there was a wide range of variation among the studied genotypes at both locations for most of traits including plant height, total number of pods plant⁻¹, number of seeds plant⁻¹, number of fertile pods plant⁻¹, dry pod yield plant⁻¹, seed weight plant⁻¹, hundred seed weight, harvest index, biological yield and seed yield ha⁻¹ while other traits showed low to fairly high range value (Table 5 and 6). At Melkassa, days to 50% flowering ranged from 36.98 to 47.69 with a mean of 41.25. Accordingly, genotype G-14 took shortest (36.98) days whereas G-10 took longest (47.69) days to attain 50% flowering than the other common bean genotypes. Nearly 5 and 66% of genotypes flowered earlier than the standard check G-63 and G-64, respectively. The longest maturity period was recorded for G-35 while, the genotype G-11 took the shortest days to mature. 47 and 72% of genotypes earlier mature than the standard check G-63 and G-64, respectively.

Twenty-four (37.5%) genotypes have matured in fewer days

Table 3: Mean square from the analysis of variance for 18 traits of 64 small seeded common bean genotypes tested at Melkassa in 2021/2022

| Traits | Mean squares | | | Error | | CV (%) | R ² (%) | Efficiency relative to RCBD (%) |
|--------|--------------------|---------------------|------------------|----------------------|---------------|--------|--------------------|---------------------------------|
| | Replication (df:2) | Block (Rep) (df:21) | Genotype (df:63) | Intra block (df:105) | RCBD (df:126) | | | |
| SCE | 217.65 | 33.17 | 185.74** | 22.03 | 23.94 | 7.53 | 85 | 108.4 |
| DF | 31.83 | 3.68 | 27.44** | 2.73 | 2.89 | 4.00 | 87 | 105.9 |
| DM | 258.06 | 10.27 | 47.76** | 5.01 | 5.89 | 2.64 | 88 | 117.6 |
| PH | 1280.49 | 90.90 | 379.65** | 61.92 | 76.86 | 10.56 | 81 | 107.9 |
| PL | 4.87 | 0.54 | 2.84** | 0.48 | 0.49 | 7.78 | 80 | 102.1 |
| PD | 1.13 | 0.29 | 0.79** | 0.26 | 0.36 | 10.39 | 68 | 102.2 |
| SCH | 655.89 | 57.04 | 244.41** | 35.47 | 39.65 | 10.53 | 83 | 110 |
| TNPPP | 119.26 | 13.49 | 54.75** | 9.57 | 10.23 | 15.87 | 80 | 106.9 |
| NFPPP | 151.52 | 10.84 | 51.61** | 10.48 | 10.54 | 17.99 | 77 | 100.6 |
| NSPP | 2.85 | 0.62 | 0.70** | 0.20 | 0.41 | 8.28 | 75 | 134.6 |
| TNSPP | 4275.6 | 340.6 | 2084.8** | 364.59 | 36.60 | 21.81 | 79 | 100 |
| SPDW | 0.14 | 0.04 | 0.09** | 0.02 | 0.022 | 10.52 | 77 | 113.9 |
| DPYPP | 109.88 | 26.1 | 93.94** | 16.61 | 18.19 | 16.90 | 79 | 109.5 |
| SWPP | 39.14 | 23.22 | 54.81** | 12.19 | 14.03 | 17.97 | 76 | 115.1 |
| HSW | 3.129115 | 2.44 | 28.82** | 1.50 | 1.66 | 5.38 | 92 | 110.6 |
| HI | 288.74 | 95.40 | 128.42** | 43.64 | 52.28 | 13.87 | 70 | 119.8 |
| BMYPH | 30146363.4 | 1231420.5 | 3993747.4** | 782177.5 | 857051.30 | 14.36 | 80 | 109.6 |
| SYPH | 13215426.22 | 711627.75 | 855919.79** | 200764 | 285887.94 | 15.39 | 82 | 142.4 |

df: Degrees of freedom; CV: Coefficient of variation; R²: Coefficient of determination; SCE: Stand count at emergence; DF: Days to 50% flowering; DM: Days to 90% maturity; PH: Plant height; PD: Pod diameter; PL: Pod length; SCH: Stand count at harvest; TNPPP: Total number of pods plant⁻¹; NFPPP: Number of fertile pod plant⁻¹; NSPP: Number of seed pod⁻¹; TNSPP: Total number of seed plant⁻¹; SPDW: Single pod dry weight; DPYPP: Dry pod yield plant⁻¹; SWPP: Seed weight plant⁻¹; HSW: Hundred seed weight; HI: Harvest index; BMYPH: Above ground biological yield ha⁻¹; SYPH: Seed yield ha⁻¹; **: Highly significant at ($p=0.01$) level of significance

than the grand mean (84.95) days from 64 genotypes to reach physiological maturity stage and these genotypes could be early maturing. Those genotypes which took shorter days to mature (early maturing genotypes) such as G-11, G-28, G-31, G-46, and G-55 can be recommendable for moisture stress area due to they need shorter rainy season and can escape moisture stress. The highest plant height was recorded by genotype G-6 (98.59 cm) followed by G-40 (93.77 cm) and G-54 (91.88 cm) while the lowest plant height was recorded in genotype G-55 (47.64 cm). A wide range was observed in number of fertile pods per plant varied from 10.05 to 29.35 with a grand mean of 17.99. The highest pods per plant (29.35 pods) were scored for genotypes G-35. A significant variation for number of seeds per plant ranged from 44.25 to 162.09 with a grand mean of 87.56. The maximum number of seeds per plant was obtained from G-35 whereas the check G-64 produced

minimum number of seeds plant⁻¹. The maximum seed weight plant⁻¹ (33.19 g plant⁻¹) was recorded from genotype G-35 whereas the minimum seed weight plant⁻¹ (10.36 g plant⁻¹) was produced for G-53.

At Kulumsa, genotype G-41 took shortest (47.40) days whereas G-58 took longest (58.96) days to attain 50% flowering. The shortest maturity period (105.69) was recorded for G-39 while, the genotype G-58 took the longest days to maturity (119.04). Yohannes et al. (2020) reported a variation of days to maturity, ranged from 75 to 92 days in common bean. Genotypes which took longer days to mature (late maturing genotypes) such as G-58, G-43, G-61, G-53, G-54 and G-36 can be recommendable for non stress/potential/mid altitude areas like Kulumsa and similar agro-ecologies. The highest plant height was scored for G-5 (96.54) whereas the lowest plant height was recorded in genotype G-39 (42.55 cm). The highest

Table 4: Mean square from the analysis of variance for 18 traits of 64 small seeded common bean genotypes tested at Kulumsa in 2021/2022

| Traits | Mean squares | | | Error | | CV (%) | R ² (%) | Efficiency relative to RCBD (%) |
|--------|--------------------|---------------------|------------------|----------------------|---------------|--------|--------------------|---------------------------------|
| | Replication (df:2) | Block (Rep) (df:21) | Genotype (df:63) | Intra block (df:105) | RCBD (df:126) | | | |
| SCE | 159.15 | 41.05 | 170.60** | 29.02 | 31.02 | 7.91 | 80 | 106.89 |
| DF | 2.90 | 1.64 | 22.26** | 1.55 | 1.56 | 2.4 | 89.9 | 100.65 |
| DM | 18.19 | 4.79 | 22.78** | 3.25 | 3.51 | 1.6 | 82 | 108.00 |
| PH | 445.54 | 136.34 | 467.59** | 64.69 | 76.6 | 10.64 | 83 | 118.41 |
| PL | 2.23 | 0.28 | 2.72** | 0.099 | 0.128 | 3.74 | 94.6 | 129.29 |
| PD | 0.96 | 0.38 | 0.68** | 0.186 | 0.219 | 14.16 | 73.1 | 117.74 |
| SCH | 28.35 | 43.80 | 192.70** | 29.09 | 31.54 | 8.36 | 81 | 108.42 |
| TNPPP | 90.78 | 14.84 | 47.11** | 8.12 | 9.24 | 14.82 | 80.2 | 113.79 |
| NFPPP | 88.26 | 17.21 | 45.18** | 8.19 | 9.69 | 15.74 | 79.7 | 118.32 |
| NSPP | 0.59 | 0.31 | 0.76** | 0.22 | 0.24 | 9.01 | 70.4 | 109.09 |
| TNSPP | 1476.27 | 735.92 | 1458.81** | 220.54 | 306.4 | 17.85 | 82.6 | 138.93 |
| SPDW | 0.02 | 0.04 | 0.25** | 0.028 | 0.029 | 11.1 | 85 | 103.57 |
| DPYPP | 71.01 | 25.13 | 48.51** | 10.43 | 12.88 | 14.7 | 77.3 | 123.49 |
| SWPP | 71.19 | 20.98 | 27.97** | 6.77 | 9.14 | 15.3 | 76.7 | 135.01 |
| HSW | 2.26 | 1.36 | 82.34** | 0.73 | 0.83 | 4.03 | 98.6 | 113.70 |
| HI | 16.41 | 13.27 | 113.05** | 16.51 | 15.97 | 8.43 | 81.1 | 96.73 |
| BMYPH | 3749084.5 | 1779683.4 | 3192655.4** | 702364.5 | 881918 | 11.3 | 76.9 | 125.56 |
| SYPH | 663306.55 | 416944.48 | 1239857.6** | 202802.2 | 238493 | 12.59 | 80.55 | 117.60 |

df: Degrees of freedom; CV: Coefficient of variation; R²: coefficient of determination; SCE: Stand count at emergence; DF: Days to 50% flowering; DM: Days to 90% maturity; PH: Plant height; PD: Pod diameter; PL: Pod length; SCH: Stand count at harvest; TNPPP: Total no. of pods plant⁻¹; NFPPP: No. of fertile pod plant⁻¹; NSPP: No. of seed pod⁻¹; TNSPP: Total no. of seed plant⁻¹; SPDW: Single pod dry weight; DPYPP: Dry pod yield plant⁻¹; SWPP: Seed weight plant⁻¹; HSW: Hundred seed weight; HI: Harvest index; BMYPH: Above ground biological yield ha⁻¹; SYPH: Seed yield ha⁻¹; **: Highly significant at ($p=0.01$) level of significance

pods plant⁻¹ (26.8 pods) were scored for genotypes G-4. The maximum number of seeds plant⁻¹ (142.60) and the highest seed weight plant⁻¹ (23.85 g plant⁻¹) was recorded from G-44. Seed yield showed a wide range of variation (1685.09 kg ha⁻¹ to 4499.61 kg ha⁻¹) with a mean value of 2911.04 kg ha⁻¹ at Melkassa and from 1369.76 kg ha⁻¹ to 4848.38 kg ha⁻¹ with a mean of 3576.82.4 kg ha⁻¹ at Kulumsa. In agreement with the present finding, Girum (2019) reported a significant difference in grain yield ha⁻¹ ranged from 1668.8 kg ha⁻¹ to 4014.7 kg ha⁻¹ with a mean of 2863.7 kg ha⁻¹ in thirty common bean genotypes. Abnet (2020) also found a wide range of variation in seed yield ha⁻¹ which ranged from 2178 kg ha⁻¹ to 5623.5 kg ha⁻¹ among forty nine common bean genotypes.

The highest yielding genotypes at Melkassa were G-39(4499.61 kg ha⁻¹) followed by G-27(4157.1 kg

ha⁻¹), G-58(3992.03 kg ha⁻¹), G-35(3868.8 kg ha⁻¹), G-29(3683.48 kg ha⁻¹), G-18(3631.18 kg ha⁻¹), G-8(3628.31 kg ha⁻¹), G-61(3595.21 kg ha⁻¹), G-37(3580.84 kg ha⁻¹) and G-33(3550.42 kg ha⁻¹). These genotypes showed from 49% to 88.9% yield advantage over the best standard check G-64. while the top high yielding genotypes at Kulumsa were G-33(4848.38 kg ha⁻¹), G-8(4730.12 kg ha⁻¹), G-4(4635.18 kg ha⁻¹), G-12(4431.71 kg ha⁻¹), G-58(4394.55 kg ha⁻¹), G-11(4365.80 kg ha⁻¹), G-45(4263.45 kg ha⁻¹), G-13(4157.19 kg ha⁻¹), G-36(4152.02 kg ha⁻¹) and G-30(4137.80 kg ha⁻¹). These genotypes also showed from 16% to 36% yield advantage over the best standard check G-64. Therefore; the above mentioned genotypes are promising which could be exploited in breeding program for further evaluation advanced to variety trial and or as a parent for improvement of yield and yield component traits. G-33, G-58 and G-8 gave higher mean grain yield consistently



Table 5: Range; mean and estimates of genetic parameters for 18 traits of small seeded common bean genotypes at Melkassa

| Traits | Range | Mean±SE | σ^2_g | σ^2_p | GCV % | PCV % | H ² B% | GA | GAM % |
|--------|-----------------|----------------|--------------|--------------|-------|-------|-------------------|--------|-------|
| SCE | 47.08-74.94 | 62.30±2.92 | 52.84 | 74.84 | 11.67 | 13.89 | 70.60 | 12.60 | 20.23 |
| DF | 36.98-47.69 | 41.25±1.03 | 8.4 | 11.15 | 7.03 | 8.09 | 75.34 | 5.19 | 12.58 |
| DM | 76.46-90.88 | 84.95±1.39 | 13.46 | 18.47 | 4.32 | 5.06 | 72.87 | 6.46 | 7.61 |
| PH | 47.64-98.59 | 74.48±4.90 | 109.68 | 171.75 | 14.06 | 17.60 | 63.86 | 17.27 | 23.18 |
| PL | 7.23-11.23 | 8.92±0.43 | 0.79 | 1.27 | 9.96 | 12.63 | 62.20 | 1.45 | 16.21 |
| PD | 3.31-6.31 | 4.89±0.32 | 0.16 | 0.42 | 8.18 | 13.25 | 38.10 | 0.51 | 10.42 |
| SCH | 38.54-71.69 | 56.55±3.71 | 72.01 | 107.54 | 15.01 | 18.34 | 66.96 | 14.33 | 25.33 |
| TNPPP | 10.95-30.99 | 19.50±1.93 | 15.2 | 24.76 | 19.99 | 25.52 | 61.39 | 6.30 | 32.32 |
| NFPPP | 10.05-29.35 | 17.99±2.02 | 13.65 | 24.12 | 20.54 | 27.30 | 56.59 | 5.73 | 31.87 |
| NSPP | 4.13-6.62 | 5.43±0.28 | 0.25 | 0.46 | 9.21 | 12.49 | 54.35 | 0.76 | 14.00 |
| TNSPP | 44.25-162.09 | 87.56±11.89 | 581.5 | 947.70 | 27.54 | 35.16 | 61.36 | 38.97 | 44.50 |
| SPDW | 0.96-1.83 | 1.33±0.09 | 0.03 | 0.04 | 11.95 | 15.92 | 56.31 | 0.25 | 18.50 |
| DPYPP | 12.90-43.21 | 24.11±2.54 | 26.11 | 42.78 | 21.19 | 27.13 | 61.03 | 8.24 | 34.16 |
| SWPP | 10.36-33.19 | 19.43±2.18 | 14.45 | 26.72 | 19.56 | 26.60 | 54.08 | 5.77 | 29.68 |
| HSW | 18.06-32.28 | 22.76±0.76 | 9.32 | 10.82 | 13.41 | 14.45 | 86.15 | 5.85 | 25.68 |
| HI | 33.76-64.88 | 47.63±4.12 | 30.5 | 73.99 | 11.59 | 18.06 | 41.22 | 7.31 | 15.36 |
| BMYPH | 3931.99-9204.89 | 6159.81±550.91 | 1072427 | 1858459 | 16.81 | 22.13 | 57.71 | 1622.9 | 26.35 |
| SYPH | 1685.09-4499.61 | 2911.04±279.11 | 225804 | 428411 | 16.32 | 22.48 | 52.71 | 711.71 | 24.45 |

SE: Standard error; σ^2_g : Genotypic variance; σ^2_p : Phenotypic variance; GCV: Genotypic coefficient of variation; PCV: Phenotypic coefficient of variation; H²B: Broad sense heritability; GA: Genetic Advance; GAM: Genetic advance as percentage of mean; SCE: Stand count at emergence; DF: Days to 50% flowering; DM: Days to 90% maturity; PH: Plant height; PD: Pod diameter; PL: Pod length; SCH: Stand count at harvest; TNPPP: Total number of pods plant⁻¹; NFPPP: No. of fertile pod plant⁻¹; NSPP: No. of seeds pod⁻¹; TNSPP: Total no. of seeds plant⁻¹; SPDW: Single pod dry weight; DPYPP: Dry pod yield plant⁻¹; SWPP: Seed weight plant⁻¹; HSW: Hundred seed weight; HI: Harvest index; BMYPH: Biological yield ha⁻¹; SYPH: Seed yield ha⁻¹

at both locations. In general, the yield performance was good at both locations. However, comparatively it was better at Kulumsa, indicating its potential for common bean production.

3.1. Phenotypic and genotypic coefficient of variation

Phenotypic coefficient of variation and genotypic coefficient of variation reveals the extent of variability present for different characters and used to measure the amount of genetic variation that exists in a given population but not the heritable portion of variability (Burton and Devane, 1953). The present study indicated that PCV value ranged from 5.06% for days to maturity to 35.16% for number of seed plant⁻¹, and GCV value ranged from 4.32% for days to maturity to 27.54% for number of seed plant⁻¹ at Melkassa (Table 5). According to Deshmukh et al. (1986), the phenotypic coefficient of variation (PCV) and the genotypic coefficient of variation (GCV) values can be categorized as low (<10%), moderate (10–20%), and high

(>20%). Based on these categories, relatively high PCV and GCV values were recorded at Melkassa for number of seeds plant⁻¹, number of fertile pods plant⁻¹, dry pod yield plant⁻¹, total number of pods plant⁻¹ and seed weight plant⁻¹ (Table 5). Traits such as number of seeds plant⁻¹, hundred seed weight, number of fertile pods plant⁻¹ and total number of pods plant⁻¹ were showed relatively high PCV and GCV at Kulumsa (Table 6). These higher PCV and GCV values specified that the genotypes in this study had a wide genetic basis and high variability among themselves with respect to these traits. This indicated that selection may be effective based on these traits and their phenotypic expression would be a good indication of genotypic potential. In agreement with this result, Alemayehu (2010) reported high PCV and GCV for number of pods plant⁻¹ and seed weight plant⁻¹ in common bean. Panchbhaya et al. (2017) observed high PCV and GCV for number of pods plant⁻¹, pod yield and seed weight plant⁻¹. Higher PCV and GCV value for number of pods plant⁻¹ was also reported by Wondwosen

Table 6: Range, mean and estimates of genetic parameters for 18 traits of Small seeded Common bean genotypes at kulumsa

| Traits | Range | Mean±SE | σ^2_g | σ^2_p | GCV % | PCV % | H ² B % | GA | GAM % |
|--------|-----------------|----------------|--------------|--------------|-------|-------|--------------------|---------|-------|
| SCE | 48.42-80.50 | 68.1±3.56 | 47.6 | 76.79 | 10.13 | 12.87 | 61.99 | 11.19 | 16.43 |
| DF | 47.4-58.96 | 51.76±0.77 | 6.94 | 8.49 | 5.09 | 5.63 | 81.74 | 4.91 | 9.48 |
| DM | 105.69-119.04 | 112.79±1.12 | 6.67 | 9.92 | 2.29 | 2.79 | 67.24 | 4.36 | 3.87 |
| PH | 42.55-96.54 | 75.59±5.01 | 119.43 | 184.2 | 14.46 | 17.95 | 64.84 | 18.13 | 23.98 |
| PL | 6.52-10.40 | 8.42±0.196 | 0.89 | 0.99 | 11.20 | 11.81 | 89.99 | 1.84 | 21.90 |
| PD | 2.08-4.67 | 3.05±0.27 | 0.171 | 0.36 | 13.56 | 19.59 | 47.90 | 0.59 | 19.33 |
| SCH | 49.04-80.65 | 64.54±3.36 | 57.16 | 86.33 | 11.71 | 14.40 | 66.21 | 12.67 | 19.64 |
| TNPPP | 11.72-27.94 | 19.23±1.77 | 13.71 | 21.85 | 19.25 | 24.31 | 62.75 | 6.04 | 31.42 |
| NFPPP | 9.03-26.8 | 18.18±1.78 | 12.37 | 20.57 | 19.35 | 24.95 | 60.14 | 5.62 | 30.90 |
| NSPP | 3.79-6.37 | 5.23±0.29 | 0.17 | 0.39 | 7.88 | 11.94 | 43.59 | 0.56 | 10.72 |
| TNSPP | 32.27-142.60 | 83.2±9.25 | 430.43 | 651.82 | 24.94 | 30.69 | 66.04 | 34.73 | 41.74 |
| SPDW | 1.11-2.68 | 1.5±0.10 | 0.078 | 0.11 | 18.62 | 21.71 | 73.58 | 0.49 | 32.90 |
| DPYPP | 10.02-31.62 | 21.99±2.01 | 12.92 | 23.43 | 16.35 | 22.01 | 55.14 | 5.50 | 25.00 |
| SWPP | 8.14-23.85 | 17±1.62 | 6.81 | 13.62 | 15.35 | 21.71 | 50 | 3.80 | 22.36 |
| HSW | 14.50-47.13 | 21.19±0.53 | 27.58 | 28.31 | 24.78 | 25.11 | 97.42 | 10.68 | 50.39 |
| HI | 29.88-59.02 | 48.2±2.53 | 30.59 | 47.1 | 11.47 | 14.24 | 64.95 | 9.18 | 19.05 |
| BMYPH | 7869.84-9725.31 | 7417.36±522.05 | 919702 | 1626433 | 12.93 | 17.19 | 56.55 | 1485.58 | 20.03 |
| SYPH | 1369.76-4848.38 | 3576.82±280.52 | 355439 | 559831 | 16.67 | 20.92 | 63.49 | 978.60 | 27.36 |

SE: Standard error; σ^2_g : Genotypic variance; σ^2_p : Phenotypic variance; GCV: Genotypic coefficient of variation; PCV: Phenotypic coefficient of variation; H²B: Broad sense heritability; GA: Genetic Advance; GAM: Genetic advance as percentage of mean; SCE: Stand count at emergence; DF: Days to 50% flowering; DM: Days to 90% maturity; PH: Plant height; PD: Pod diameter; PL: Pod length; SCH: Stand count at harvest; TNPPP: Total no. of pods plant⁻¹; NFPPP: No. of fertile pod plant⁻¹; NSPP: Number of seeds pod⁻¹; TNSPP: Total number of seeds plant⁻¹; SPDW: Single pod dry weight ; DPYPP: Dry pod yield plant⁻¹; SWPP: Seed weight plant⁻¹; HSW :Hundred seed weight; HI: Harvest index; BMYPH: Biological yield ha⁻¹; SYPH: Seed yield ha⁻¹

and Abebe, 2017; Ghimire and Mandal, 2019). Similarly, higher PCV for number of seeds per plant was reported by many researchers (Yonas, 2017; Abnet, 2020; Temesgen, 2020) but the authors found moderate GCV contradicting with the present result. The difference among the present result and the previous studies for GCV with respect to this trait may be differences in genotype. At Melkassa, seed yield and biological yield ha⁻¹ were recorded high PCV and moderate GCV values. Seed weight plant⁻¹, single pod dry weight, dry pod yield plant⁻¹ and Seed yield ha⁻¹ had high PCV and moderate GCV values at kulumsa. In line with these finding different researchers reported high PCV and moderate GCV for seed yield ha⁻¹ (Ejigu et al., 2018) and for biological yield ha⁻¹ (Aziza, 2019; Kefelegn et al., 2020). Moderate PCV and GCV (10–20%) values were observed in harvest index, stand count at emergence, plant height and stand count at harvest at both locations (Table 5 and 6). These moderate values indicated the existence of enough genetic variation on the studied genotypes to perform

selection for improvement. Similar results have been noted by different authors such as Alemayehu (2010) and Wondwosen and Abebe (2017) for hundred seed weight, Aziza (2019) for harvest index, Bagheri et al. (2017) and Ghimire and Mandal (2019) for plant height, Panchbhayia et al. (2017) and Jhanavi et al. (2018) for single pod weight.

However, days to 50% flowering and days to 90% maturity showed lower GCV and PCV values. The low value of this variation indicates that selection is not effective for these traits, because of the narrow range of variations even though it showed less influence of environmental effect on the expression of these traits at both locations. In order to improve those traits there is a requirement of creation of genetic variation through hybridization and or induced mutagenesis followed by selection. Yonas (2017) and Ejigu et al. (2018) reported low GCV and PCV for days to 50% flowering and days to 90% maturity in common bean genotypes. In addition, Abnet (2020) also observed low PCV and GCV values for days to 50% flowering and days

to maturity in forty nine common bean genotypes similar to the present result.

Moderate PCV and low GCV values were recorded by pod length, and pod diameter at Melkassa and number of seeds per pod at both Melkassa and Kulumsa. This would indicate the presence of environmental influence on the phenotypic expression of these traits and low range of genetic variation. Hence, these traits also lower responsive for selection. Aziza (2019) reported moderate PCV and low GCV for pod length and pod diameter. Moderate PCV and low GCV estimate of number of seeds pod⁻¹ was reported by Yonas (2017) and Ejigu et al. (2018) similar to the current finding. The lowest PCV and GCV (2.79, 2.29%) was recorded for days to 90% maturity and the highest (30.69, 24.94%) for number of seeds plant⁻¹ at Kulumsa respectively (Table 6).

Similar GCV and PCV patterns were observed in both locations for number of seeds plant⁻¹, number of fertile pods plant⁻¹, total number of pods plant⁻¹, seed yield ha⁻¹, harvest index, stand count at emergence, plant height, stand count at harvest, number of seeds pod⁻¹, days to 50% flowering and days to 90% maturity. Generally, in the present study, the phenotypic coefficient of variation (PCV) was higher than genotypic coefficient of variation (GCV) at both locations for all traits although the differences were not large for most of traits. This would be due to the fact the variation at the phenotypic level was due to the effect of genotypes and influence of environment. PCV values for most of the traits at Melkassa were higher than that observed at Kulumsa. This could be due to high environmental influence for phenotypic expression of those traits at Melkassa. Consistently higher PCV and GCV values were observed in both environments for number of seeds plant⁻¹, number of fertile pods plant⁻¹ and total number of pods plant⁻¹.

3.2. Estimates of heritability (H²B) in a broad sense

Broad sense heritability gives an idea about portion of observed variability attributable to genetic difference. The broad sense heritability values were ranged from moderate to high at both locations (Table 5 and 6). It was ranged from 38.10% for pod diameter to 86.15% for hundred seed weight at Melkassa (Table 4) and from 43.59% for number of seeds pod⁻¹ to 97.42% for hundred seed weight at Kulumsa (Table 5). Heritability estimates in broad sense was categorized as high (>60%), medium (30–60%) and low (<30%) (Dabholkar, 1992). Based on this classification, most of the traits have shown consistently high heritability values in both locations such as stand count at emergence, days to 50% flowering, days to 90% maturity, plant height, pod length, stand count at harvest, total number of pods plant⁻¹, number of seeds plant⁻¹ and hundred seed weight (Table 5 and 6). At Kulumsa, pod length, number of fertile pods plant⁻¹, single pod dry weight, harvest index and seed yield

per hectare also scored high heritability. Higher heritability estimates for those traits indicated that the variation observed was mainly under genetic control and was less influenced by environment. In supporting of this finding different researcher reported high broad sense heritability for hundred seed weight, pods per plant, days to maturity and days to flowering (Alemayehu, 2010; Panchbhaya et al., 2017; Wondwosen and Abebe, 2017).

Ejigu et al. (2018) and Temesgen (2020) obtained high heritability for seeds plant⁻¹ and plant height, respectively. Kefelegn et al. (2020) also found high heritability values for pod plant⁻¹, pod length, plant height, days to 50% flowering, days to maturity and hundred seed weight in their study in common bean which support the present findings.

Whereas, consistently moderate heritability values (30–60%) was recorded for pod diameter, number of seeds pod⁻¹, seed weight plant⁻¹ and biological yield ha⁻¹ at both locations as indicated in (Table 5 and 6). Number of fertile pods plant⁻¹, single pod dry weight, seed weight plant⁻¹, harvest index and seed yield ha⁻¹ had a moderate heritability at Melkassa. Aziza (2019) observed moderate heritability for traits like number of seeds pod⁻¹ (44.7%), pod diameter (43.8%), biological yield (54.6%) and seed yield (49.9%) in common bean genotypes in agreement with the present result. However, the author found moderate heritability (30–60) estimate for number of seeds plant⁻¹ (57.3%) and hundred seed weight (46.2%) opposing to the current heritability estimates. Kefelegn et al. (2020) also reported high heritability values for seed pod⁻¹ (71%) and biological yield (73%) in their study in common bean contradicted with the present findings. The difference among the present result and the aforementioned authors may be differences in genotype and environment often bring about differences in the results of different studies.

3.3. Estimates of genetic advance as percent of mean

Genetic advance as percent of mean (GAM) ranged from low to high at both locations (Table 5 and 6). It was ranged from 7.61% for days to maturity to 44.50% for number of seeds per plant at melkassa and from 3.87% for days to maturity to 50.39% for hundred seed weight at Kulumsa. According to Johnson et al. (1955) genetic advances as a percentage of the mean (GAM) are classified high (>20%), moderate (10–20%) and low (<10%). Based on this delineation, consistently high GAM values were observed in both locations for most of traits including plant height, total number of pods plant⁻¹, number of fertile pods plant⁻¹, number of seeds plant⁻¹, seed weight plant⁻¹, dry pod yield plant⁻¹, hundred seed weight, biological yield and seed yield ha⁻¹. A result of high GAM indicated the maximum control of characters by additive gene action and the high possibility of using this trait for genetic improvement through

selection. In harmony with present finding, high GAM for number of seeds plant⁻¹, number of pods plant⁻¹, plant height and seed yield has been reported by Anunda et al. (2019), Aziza (2019) and Kefelegn et al. (2020). However, days to 90% maturity showed lower GAM values. In addition, days to 50% flowering also showed low GAM in Kulumsa. This implies selection of genotype based on these traits will not make any improvement in new population. Similarly, low GAM for days to maturity was reported by other researchers (Yonas, 2017; Ejigu et al., 2018; Abnet, 2020).

Since high heritability does not always indicate a high genetic gain, heritability with genetic advance, considered together, should be used in predicting the ultimate effect of selecting superior varieties (Johnson et al., 1955). Accordingly, in the present study, most of the traits coupled moderate to high heritability with moderate to high genetic advance as a percent of mean except days to 90% maturity (at both locations) and days to 50% flowering (at Kulumsa) coupled with high heritability and low genetic advance indicating the presence of non-additive gene action and hence, heterosis breeding may be recommended for the improvement of these traits than selection. The present result is in agreement with Anunda et al. (2019) who reported high heritability coupled with high genetic advance for plant height, number of pods plant⁻¹, number of seeds plant⁻¹ and hundred seed weight. This indicating most likely the heritability is due to additive gene effects and selection may be effective for further improvement. Similarly, high heritability coupled with low genetic advance for days to maturity was also reported by Yonas (2017).

In addition, estimating genotypic coefficient of variation along with heritability plus genetic advance as a percent of mean are crucial to improve traits of interest by understanding the type of gene action involved in the expression of traits especially for polygenic traits (Anunda et al., 2019) and thus to provide better information and to reach more concrete conclusion than single parameters alone (Denton and Nwangburuka, 2011). Based on this fact, high GCV with moderate to high heritability coupled with high GAM were observed for number of fertile pods plant⁻¹, number of seeds plant⁻¹ and total number of pods plant⁻¹ at both locations. Dry pod yield and seed weight per plant at Melkassa and hundred seed weight at Kulumsa also showed high GCV with moderate to high heritability coupled with high GAM values in the present study which indicates that the traits were simply inherited in nature and possessed additive gene effects. In agreement to the present result, Topwal and Gaur (2016) and Bagheri et al. (2017) found higher GCV with high heritability coupled with high GAM for number of pods plant⁻¹, number of seeds plant⁻¹ and pod weight plant⁻¹ respectively.

Traits such as stand count at emergence, plant height, stand count at harvest, single pod dry weight, above ground biological yield, seed yield ha⁻¹ and harvest index exhibited moderate GCV coupled with moderate to high heritability and Genetic advance as a present of mean even though they don't showed similar trend of patterns in both locations. It indicates that the phenotype of an individual in the current population is a good indicator of the genotypes, or it may mean that most of the variation in this trait observed in the present population is caused by variation in genotypes. This reflected the involvement of additive gene action in the expression of these traits.

4. CONCLUSION

The phenotypic coefficient of variation was higher than genotypic coefficient of variation; indicated the variation at the phenotypic level was due to the effect of genotypes and influence of environment. Most of traits exhibited moderate to high GCV, heritability and GAM association, implying the importance of those traits for improving yield and associated traits. The present study generally indicated considerable genetic variability among the genotypes. Thus, there is enormous opportunity for improving yield and attributing traits through direct selection and or hybridisation.

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6. REFERENCES

- Abnet, D., 2020. Genetic variability and association of traits in small seeded common bean (*Phaseolus vulgaris* L.) genotypes in Mecha District, Amhara Region. An MSc Thesis Presented to the School of Graduate Studies of Bahirdar University.
- Alemayehu, B., 2010. Genetic variation for grain yield and water absorption in common bean (*Phaseolus vulgaris* L.). African Journal of Food Science and Technology 1(6), 128–131.
- Anonymous, 2019. Agricultural sample survey. Report on Area and Production of Major Crops, Volume I. Addis Ababa, Pp. 14.
- Anonymous, 2017. Animal and plant health regulatory directorate. Crop Genotype Register Issue No. 17, June 2017, Addis Ababa, Ethiopia.
- Anonymous, 2016. Common bean (*Phaseolus vulgaris*),

- in safety assessment of transgenic organisms in the environment, Volume 6: OECD Consensus Documents, OECD Publishing, Paris.
- Anonymous, 2020. Statistical database of agricultural production. Rome, Italy.
- Anunda, H.N., Nyaboga, E.N., Amugune, N.O., 2019. Evaluation of genetic variability, heritability, genetic advance and correlation for agronomic and yield components in common bean landraces from South Western Kenya. *Journal of Plant Breeding and Crop Science* 11(5), 144–157.
- Aziza, A., 2019. Genetic variability and trait associations of common bean (*Phaseolus vulgaris* L.) genotypes in Northeastern Amhara. An MSc Thesis Presented to the School of Graduate Studies of Bahrdar University, Ethiopia, 94p.
- Bagheri, M., Kahrizi, D., Zebajadi, A., 2017. Study on genetic variation and morpho-phenologic traits in common bean (*Phaseolus vulgaris* L.). *Biharean Biologist* 11(1), 43–47.
- Berhanu, A., Kassaye, N., Tigist, S., Kidane, T., Dagmawit, T., Rubyogo, J.C., Clare, M.M., 2018. Progress of common bean breeding and genetics research in Ethiopia. *Ethiopian Journal of Crop Science* 6(3), 115–128.
- Broughton, W.J., Hernandez, G., Blair, M.W., Beebe, S.E., Gepts, P., Vanderleyden, J., 2003. Beans (*Phaseolus* Spp.) model food legumes. *Plant and Soil* 252, 55–128.
- Burton, W.G., Devane, E.H., 1953. Estimating heritability in tall fescue from replicated clonal material. *Agronomy Journal* 45, 478–481.
- Dabholkar, A.R., 1992. Elements of biometrical genetics, Concept Pub. Co., New Delhi. 431p.
- Denton, O.A., Nwangburuka, C.C., 2011. Heritability, genetic advance and character association in six yield related characters of *Solanum Anguivi*. *Asian Journal of Agrecultural Research* 5, 201–207.
- Deshmukh, S.N., Basu, M.S., Reddy, P.S., 1986. Genetic variability, character association and path coefficient analysis of quantitative traits in virginia bunch varieties of groundnut. *Indian Journal of Agricultural Sciences* 56, 515–518.
- Ejigu, E., Wassu, M., Berhanu, A., 2018. Genetic variability, heritability and expected genetic advance of yield and yield related traits in common bean genotypes (*Phaseolus vulgaris* L.) at Abaya and Yabello. *African Journal of Biotechnology* 17(31), 973–980.
- Ephrem, T.L., 2016. Review of haricot bean value chain in Ethiopia. *International Journal of African and Asian Studies* 24, 65–72.
- Falconer, D.S., Mackay, T.F.C., 1996. Introduction to quantitative genetics. 4th Eds, Longman Group Limited, Malaysia. 464p.
- Frehiwot, M., 2010. Profile of haricot bean production, supply, demand and marketing issues in Ethiopia, Ethiopia Commodity Exchange Authority, Addis Ababa, Ethiopia, 50pp.
- Ghimire, N.H., Mandal, H.N., 2019. Genetic variability, heritability and genetic advance of common bean (*Phaseolus vulgaris* L.) genotypes at mountain environment of Nepal. *International Journal of Advanced Research and Biological Science* 6(10), 46–56.
- Girum, K., 2019. Genetic gain in grain yield and yield-associated traits from common bean (*Phaseolus vulgaris* L.) breeding In Ethiopia. An MSc Thesis Presented to the School of Graduate Studies of Hawasa University.
- Jhanavi, D.R., Patil, H.B., Justin, P., Hadimani, R.H.P., Mulla, S.W.R., Sarvamangala, C., 2018. Genetic variability, heritability and genetic advance studies in french bean (*Phaseolus vulgaris* L.) genotypes. *Indian Journal of Agricultural Research* 52(2), 162–166.
- Johnson, H.W., Robinson, H.F., Comstock, R.E., 1955. Estimates of genetic and environmental variability in soya bean. *Agronomy Journal* 47, 314–318.
- Kefelegn, N., Frew, M., Yigzaw, D., 2020. Genetic advancement and variability of released common bean (*Phaseolus vulgaris* L.) varieties from 1974 – 2009 GC in Ethiopia. *Advances in Agriculture Volume* 2020, Article ID 1315436, 7 pages.
- Ketema, H., Thangavel, S., 2016. Evaluation of common bean cultivars and fungicide spray frequency for the management of anthracnose (*Colletotrichum Lindemuthianum*) in ambo, West Shewa Zone, Ethiopia. *Journal of Biology, Agriculture and Healthcare* 6(19), 68–80.
- Panchbhaiya, A., Singh, D.K., Jatav, V., Mallesh, S., Verma, P., 2017. Studies on variability, heritability and genetic advance for yield and yield contributing characters in french bean (*Phaseolus vulgaris* L.) germplasm under Tarai Region of Uttarakh. *Journal of Applied and Natural Science* 9(4), 1926–1930.
- Temesgen, A., 2020. Genetic variability and association of traits in indeterminate common bean (*Phaseolus vulgaris* L.) genotypes in finoteselam, West Gojam, Ethiopia. An MSc Thesis Presented to the School of Graduate Studies of Bahirdar University.
- Topwal, M., Gaur, G., 2016. Studies on genetic variability in different genotypes of french bean (*Phaseolus*



- vulgaris* L.). International Journal of Life Science and Scientific Research 2(3), 219–221.
- Wondwosen, W., Abebe, B., 2017. Genetic variability, heritability and genetic advance of some haricot bean (*Phaseolus vulgaris* L.) varieties at bench-maji zone, Southwest Ethiopia. Asian Journal of Crop Science 9(4), 133–140.
- Yohannes, S., Gobeze, L., Mesfin, K.G., 2020. Performance evaluation of common bean (*Phaseolus vulgaris* L.) genotypes for yield and related traits at areka, Southern Ethiopia. Advances in Agriculture Volume 2020, Article ID 1497530, 8 pages.
- Yonas, M.G., 2017. Genetic diversity in speckled bean (*Phaseolus vulgaris* L.) genotypes in Ethiopia. Journal of Plant Breeding and Crop Science 9(11), 200–207.