

Effect of Bt Transgene on AM Fungi Infection and Yield Attributes of Cotton (*Gossypium hirsutum* L.)

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Abstract

A pot experiment was carried out in 2010 during wet season (July to December) at the Institute of Agricultural Sciences, Banaras Hindu University to evaluate the extent of root infection by AM (Arbuscular Mycorrhiza) fungi and yield attributing characteristics of cotton under both Bt (*Bacillus thuringiensis*) and non-Bt systems under varied soil types. It included three different soil orders i.e. *Entisol*, *Inceptisol* and *Alfisol*. Bt cotton (cv. NCS-138) and its non-transgenic isoline (cv. NCS-138) were grown until maturity. From the results it was evident that among the three different soils, red soil exhibited higher root infection in comparison to alluvial and black soil. Per cent root infection decreased significantly to the extent of 10-13% in Bt crop as compared to non-Bt counterpart and 9.5-15% during the three growth stages. During the intermediate stages of crop growth, there were significant differences in growth parameters and yield attributes between Bt and the non-Bt isoline. It was also observed that the alluvial soil produced higher shoot biomass compared to black and red soil during initial stages of crop growth. But during the intermediate stages of crop cycle, the growth picked up in black soil. In the final stage, alluvial soil produced higher biomass as compared to the black and red soil. At the same time, Bt-cotton plants showed comparatively more root length than non-Bt crop over the entire crop growth stages.

1. Introduction

Cotton (*Gossypium hirsutum* L.) is the most important fiber crop in India which plays a key role in economic and social development. India is a leading cotton producing country, and the area under Bt cotton cultivation was 11 m ha during 2013 (James, 2013) but until now there is hardly any information (especially quantitative data) generated from India about the impact of Bt cotton on soil microbiological processes (Beura and Rakshit, 2011). Although there is large-scale adoption of Bt-cotton by the farmers because of immediate financial gain, there is concern that transgenic Bt-crops (which release Bt-toxins into the environment) affect yield and microbial parameters in the agro-ecosystem. These toxins are produced in every major part of Bt-cotton plants (Dong and Li, 2007). Thus transgenic plants have the potential to modify the rhizosphere chemistry by altering plant residue quality (Dunsfield and Germida, 2004; Motavalli et al., 2004; O'callaghan et al.,

2005). Any change in the quality of rhizosphere exudates can modify the soil biota composition as well as their activity (Stotzky, 2004; Patra et al., 2006) and may produce changes in microflora and microfauna (Wei et al., 2006; Griffiths et al., 2006). Bt cotton may differ from its non-Bt counterpart in some agronomic and physiological traits, but lint yield remains unchanged due to the manipulation of boll numbers, boll weight and lint percentage under insect-free condition (Dong et al., 2006); Blanche et al. (2006) found that cultivars containing the Bollgard gene (the gene for the insecticidal protein from *Bacillus thuringiensis* spp. kurstaki; Monsanto Co.) yielded more than the conventional cultivars under optimal growth condition. With the above backdrop, our research was designed to evaluate yield attributes of cotton under both Bt and non-Bt systems under different soil types.

2. Materials and Methods

A pot experiment was carried out in 2010 during wet season



(July to December) with three different soil types at the Institute of Agricultural sciences, Banaras Hindu University, Varanasi (25°19' 60 N Latitude and 83°0' 0 E Longitude). Bt cotton (cv. NCS-138) and its non-transgenic isolate (cv. NCS-138) were grown until maturity. A no-crop reference pot was maintained with three replications for all the three soil types. The experiment was laid out in a factorial complete randomized block design with three replications.

2.1. Analysis of soil samples

Surface samples (0-15 cm) from cultivated soils of three orders i.e. *Entisol*, *Inceptisol* and *Alfisol* were collected from different geographical locations in the Varanasi and Mirzapur districts of Uttar Pradesh, India. The air-dried soils samples used in the pot experiment were ground and sieved through a 2 mm sieve, and then stored in plastic bags. An initial soil sample was analyzed for different physico-chemical parameters following the standard protocol. The pH and electrical conductivity of the soil was determined by Glass electrode pH meter and Conductivity meter (Jackson, 1973) respectively. The textural class was determined by (Bouyoucos, 1962). Rhizospheric soil samples were analyzed for mineral-N by (Subbiah and Asija, 1956), available P by (Olsen et al., 1954), available-K by (Hanway and Heidel, 1952), organic carbon by (Walkley and Black, 1934) and available cationic micronutrients by (Lindsay and Norvell, 1978) after the harvesting of cotton crop.

Root infection was assessed on a representative root sample taken from each plot at harvest. At harvest roots of 15 cm were taken from plants evenly distributed in each plot. Mycorrhiza infection of each plant was determined by estimating the percent of root segments colonised with AM with the method as described by (Bierman and Linderman, 1981).

Percent root infection was obtained as follows:

$$\% \text{ Root infection} = \frac{\text{Number of root segments infected with AM}}{\text{Total number of segments}} \times 100$$

The root length was determined by using conventional technique while the shoot and root biomass was determined after drying the roots for 24 hrs at 60-70 °C in an oven. At maturity, the total number of bolls and the boll size (weight) were recorded.

3. Results and Discussion

3.1. AM fungi infection in roots of cotton plant

The data in the Table 2 revealed that Bt-cotton affected AM fungi during the complete life cycle. Per cent root infection decreased significantly to the extent of 10-13% in Bt crop as compared to non-Bt counterpart and 9.5-15% during the three growth stages. Although AM colonization of cotton

normally progresses quickly during the first few weeks of growth reaching a plateau at 100 DAS with 45-60% of root length colonized by arbuscules. The pattern of development of mycorrhizal colonization in our assessment was a typical sigmoid pattern of logistic growth over 20 weeks of assessment. Among the three soils, red soil exhibited higher root infection as compared to alluvial and black soil, which can be explained by the differences in available P value. AM fungi colonized both the Bt and non-Bt cotton cultivars equally, providing firm evidence that both the Bt and non-Bt cotton cultivars were equally capable of establishing mycorrhizal symbiosis (Glandorf et al., 1997). The lack of differences in colonization between Bt and conventional cotton that we observed corroborates with that reported for GM and conventional soybean (Powell et al., 2007) but is in contrast to reports of differential mycorrhizal colonization of GM corn (Turrini et al., 2004; Castaldini et al., 2005).

3.2. Yield attributes of cotton

The pot experiment also revealed that Bt-cotton was able to resist insect infestation resulting in good boll retention and higher yields compared to non-Bt cotton crop. Thus, less chemical spray was required to control insect attack in Bt-cotton crop. Seed cotton yield in Bt-cotton crop ranged from 26.85 to 83.92 g pot⁻¹ in alluvial soil while 13.6-46.7 g pot⁻¹ in red soil. So, in the present study, an average yield increase

Table 1: Physico-chemical characterization of the soils

Parameters	Values		
Physical	Red soil	Black soil	Alluvial soil
Bulk density (Mg m ⁻³)	1.38	1.51	1.43
Particle density (Mg m ⁻³)	2.51	2.60	2.56
Water holding capacity (%)	39.4	45.40	41.6
Sand (%)	46.00	11.7	48.78
Silt (%)	32.85	52.7	30.48
Clay (%)	21.15	35.6	20.44
Soil texture	Silty clay loam	Clayey	Sandy loam
Electro-chemical and chemical			
pH _w (1:2.5)	6.3	7.5	7.1
Electrical conductivity (d Sm ⁻¹)	0.32	0.61	0.45
CEC {Cmol (p+) kg ⁻¹ }	18.25	31.85	19.55
Organic carbon (%)	0.34	0.42	0.38
Available nitrogen (kg ha ⁻¹)	176	238	232
Available phosphorus (kg ha ⁻¹)	8	14	18
Available potassium (kg ha ⁻¹)	110	165	148
Zn (ppm)	3.64	2.7	2.86
Ca (meq 100g ⁻¹)	3.1	32.5	10.2

of 30.6% in Bt-cotton crop compared to non Bt-cotton crop was observed due to effective control of insects. The increase seed cotton yield in this cultivar may be attributed to better fruiting efficiency, efficient source-sink relationship, in-built resistance to bollworms, more number of picked bolls plant⁻¹ and finally higher seed cotton yield. Such yield advantage of Bt-cotton hybrids over non-Bt cotton hybrids were also observed by (Halemani et al., 2004; Hallikeri et al., 2004; Srinivasulu et al., 2006).

Table 2: Root infection (%) by AM fungi of Cotton at different growth stages

DAS	Cultivar (C)	Soil types (S)			
		S ₁	S ₂	S ₃	Mean
50	Non-Bt (V ₁)	35.3	19.66	21.33	24.43
	Bt (V ₂)	31	16.33	18.33	21.88
	Mean	33.15	17.99	19.83	
	SEm±	C=0.32, S= 0.393, C×S=0.556			
	CD (p=0.05)	C=1.01, S=1.24, C×S=1.754			
100	Non-Bt (V ₁)	60.3	48.33	56	54.87
	Bt (V ₂)	54.33	44.66	51	49.99
	Mean	57.31	46.49	53.5	
	SEm±	C=0.475, S=0.58, C×S=0.826			
	CD (p=0.05)	C=1.50, S=1.82, C×S=2.6			
150	Non-Bt (V ₁)	24.66	20.00	20.66	21.77
	Bt (V ₂)	22.66	17.33	17.66	19.21
	Mean	23.66	18.66	19.16	
	SEm±	C=0.173, S=0.212, C×S=0.301			
	CD (p=0.05)	C=0.545, S=0.668, C×S=0.95			

(S₁: Red soil; S₂: Black soil; S₃: Alluvial soil; V₁: Non-Bt cultivar; V₂: Bt cultivar; V₃: No crop; DAS: Days after sowing).

3.3. Root characteristics of cotton plant

Root characteristics are the important factors that influence the rhizosphere biochemistry and the transformation of nutrients. One of the major avenues for release of Bt-toxin into the soil is through root exudates of Bt-crops. Similar trends were also documented by Benedict and Ring (2004). From the data in Table 4 it is evident that alluvial soil produced higher shoot biomass compared to black and red soil during initial stages of crop growth. But during the intermediate stages of life cycle, the growth picked up in black soil. In the final stage, alluvial soil produced higher biomass compared to black and red soil. Root biomass did not exhibit significant difference among Bt and non-Bt crop (Table 5). At the initial growth stage (50 DAS), root volume was similar for both the crops, but during later stage (100 and 150 DAS) root volume showed significant differences among Bt and non-Bt cultivars. Root length did not vary much between Bt and non-Bt-cotton crops at initial growth stages (50 DAS). While at 100 DAS, there were significant differences in root length among Bt and the non-Bt isolate. In general Bt-cotton plants showed comparatively more root length than non-Bt crop over the entire crop growth stages. Among the different soil types, alluvial soil produced higher root volume and length compared to black and red soil. The results are in close proximity with the result reported by other investigators (Hebbbar et al., 2007 and Prakash et al., 2008).

Plant height did not vary much between Bt and non-Bt cotton crops at initial stage of crop growth (50 DAS). But during the intermediate stages, plant height varied significantly among Bt and non-Bt isolate (Table 5). It is further evident that alluvial soil produced higher shoot biomass compared to black and red soil during initial stages. But during the intermediate stages of life cycle, the growth picked up in black soil. In the final

Table 3: Yield attributes at different growth stages

	No. of bolls plant ⁻¹						Bolls weight (g plant ⁻¹)					
	100 DAS			150 DAS			100 DAS			150 DAS		
	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
S ₁	3	5	4	9	12	10.5	10.46	16.74	13.6	38.3	55.1	46.7
S ₂	7	8	7.5	18	21	19.5	22.66	28.26	25.43	74.61	97.02	85.81
S ₃	7	9	8	16	19	17.5	23.93	29.78	26.85	74.75	93.09	83.92
Mean	5.66	7.33		14.33	17.33		19.01	24.92		62.55	81.73	
SEm±												
S		0.273			0.608			0.393			0.58	
V		0.223			0.495			0.32			0.475	
S×V		0.387			0.86			0.556			0.826	
CD (p=0.05)												
S		0.862			1.91			1.24			1.82	
V		0.704			1.56			1.01			1.50	
S×V		1.22			2.921			1.754			2.6	



Table 4: Shoot parameters at different growth stages

	Plant height (cm)						Dry weight in (g pot ⁻¹)					
	50 DAS			100 DAS			150 DAS			50 DAS		
	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
S ₁	53	45.66	49.33	59.33	65	62.16	61.8	69	65.4	26.43	29.8	28.12
S ₂	53.66	52.66	53.16	69.33	100.2	84.76	73.6	104.7	89.15	26.33	26.33	26.33
S ₃	67	66.33	66.66	167.6	147.3	104.9	171.6	159.3	165.45	44.06	45.66	44.86
Mean	57.88	54.88		98.75	104.16		102.3	111		32.80	33.93	
SEm±												
S		0.315			0.212			0.118			0.77	
V		0.23			0.173			0.169			0.63	
S×V		0.584			0.3			0.45			1.09	
CD (<i>p</i> =0.05)												
S		0.834			0.668			0.70			2.426	
V		0.680			0.545			0.49			1.985	
S×V		1.79			0.945			0.98			3.44	

Table 5: Root parameters of cotton at different growth stages

	Root length plant ⁻¹ (m)						Root biomass (g)					
	50 DAS			100 DAS			150 DAS			50 DAS		
	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
S ₁	7.15	11.03	9.09	19.14	27.27	23.20	24.4	34.54	29.47	14.1	19.96	12.03
S ₂	11.96	14.4	13.18	38.15	41.82	39.98	37.17	46.38	41.77	20.7	23.86	22.28
S ₃	18.27	14.14	16.20	39.66	46.88	43.27	43.42	47.87	45.64	21.62	25.96	23.79
Mean	12.46	13.19		32.31	38.65		34.99	42.93		18.8	23.26	
SEm±												
S		0.456			0.391			0.65			0.151	
V		0.371			0.31			0.39			0.122	
S×V		0.644			1.101			2.34			0.214	
CD (<i>p</i> =0.05)												
S		1.43			1.24			1.63			0.337	
V		1.17			0.53			0.69			0.385	
S×V		2.032			2.1			3.65			0.675	



stage, alluvial soil produced higher biomass compared to black and red soil. Similar improvement in yield attributes and seed cotton yield due to insecticidal pest management was reported earlier by (Bhosale et al., 2004).

4. Conclusion

There were hardly any significant differences in essential ecosystem functions such as AM fungi infection and yield attributes among Bt and non-Bt isolate at any stages of crop growth as well as after harvest. Results of the present study did not provide any firm evidence so that we can conclude both Bt and non-Bt cotton cultivars were equally capable of establishing mycorrhizal symbiosis.

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