# Cypermethrin and Methomyl Resistance in Helicoverpa armigera (Hubner)

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#### **Abstract**

The *H. armigera* larvae of the Mahaboobnagar district recorded a LD<sub>sn</sub> of 29.125 μg larva<sup>-1</sup> and 59.609 μg larva<sup>-1</sup> at LD<sub>sn</sub> for cypermethrin. The LD<sub>so</sub> and LD<sub>so</sub> values of cypermethrin for Raichur population of *H armigera* was 32.481 and 38.172 µg larva<sup>-1</sup>, respectively. Toxicity of cypermethrin to Nagpur population of H. armigera showed that the LD<sub>so</sub> and LD<sub>so</sub> values were 20.069 and 54.708 µg larva-1, respectively, The chi-square test revealed that the population used in the study was homogenous (\*p<0.05%). The H. armigera larvae of the Mahaboobnagar district recorded a LD<sub>so</sub> of 3.651 µg larva<sup>-1</sup> and 10.287 µg larva<sup>-1</sup> at LD<sub>so</sub> for methomyl. The LD<sub>so</sub> and LD<sub>so</sub> values of methomyl for Raichur population of H armigera was 3.630 and 10.417 µg larva<sup>-1</sup>, respectively, while Toxicity of methomyl to Nagpur population of H. armigera showed that the LD<sub>so</sub> and LD<sub>so</sub> values were 2.652 and 7.214 μg larva<sup>-1</sup>, respectively, when the chi-square test revealed that the population used in the study was homogenous (\*p<0.05%). From, the results obtained we can conclude that the continuous application of same chemistries across the generations increases the resistance from F, to F<sub>3</sub>. Alternating with the new chemistries results in no cross resistance development as it was observed in all the three populations.

Keywords: Helicoverpa armigera, resistance, cypermethrin, methomyl

#### 1. Introduction

The bollworm, Helicoverpa armigera Hübner (Lepidoptera: Noctuidae) is a polyphagous pest of worldwide occurrence inflicting annual crop damage in India worth US \$1 billion. In India this insect occurs as a major pest in many economically important crops, including cotton, pigeonpea, chickpea, tomato, okra, blackgram, maize, sorghum and many other crops, inflicting substantial crop losses every year (Reed and Pawar, 1982; Manjunath, 1990). Understanding the genetic variation among the H. armigera populations occurring on host plants has become essential to understand the variation in their susceptibility to different insecticides. The ability of insect species to thrive on diverse host plants is an adaptive advantage for their better survival in the ecosystem. H. armigera is also characterized by its high mobility and fecundity and it has shown great capacity to develop resistance to synthetic insecticides used in its management (Armes et al., 1996; Kranthi et al., 1997; Ramasubramaniam and Regupathy, 2004b). The versatility of this species may be due to the presence of a strong genetic variability governing the behavior of *H.armigera* making it a serious pest on several crops (Zhou et al., 2000).

The occurrence of insecticide resistant strains can be

reduced or delayed by reducing the selection pressure, by using alternate insecticides with novel mode of action. The pyrethroids and organophosphorus combination insecticides were found to be effective against the resistant insect pest population of H. armigera (Martin et al., 2003). To study the resistance levels in the field population present study with cypermethrin and methomyl was conducted.

### 2. Materials and Methods

2.1. Determination of the insecticide resistance in H. armigera

### 2.1.1. Collection of H. armigera

The larvae of *H. armigera* were collected on red gram, cotton and bengal gram crops during October, 2006 from Raichur, December-2006 and January, 2007 from Mahaboobnagar and February 2010 from Nagpur for conducting the investigation. The work was carried out in the Department of Entomology, College of Agriculture, Rajendranagar, Hyderabad, Andhra Pradesh

### 2.1.2. Test insecticides

The degree of resistance acquired by H. armigera to test insecticides viz., cypermethrin representing synthetic pyrethroids and methomyl representing carbamates group of insecticides (Table 1) were tested.



Table 1: Insecticides used for the determination of insecticide resistance in <i>H. armigera</i>									
Common name	Formulation	Trade name	Chemical name	Source of supply					
Methomyl	40 SP	Lannate	S-methylN-(methyl carbamoxyloxy) Thioacetimidate	M/S Dupont Chemical (India) Ltd, Mumbai-400076					
Cyper methrin	10 EC	Cypra	(RS)-a-cyno-3-phenoxybenzlyl-(1RS)-cis,trans-3-(2,2 dichlorovinyl) -2,2-dimethyl cyclopropanecarboxylate	M/S Hyderabad chemicals supplies Ltd, A-24/25, APIE, Balanagar, Hyder- abad-500 037					

### 2.1.3. Test insect population

The larvae collected from Mahaboobnagar, Raichur and Nagpur were reared separately in the laboratory to obtain pupae. Male, female pupae were separated and kept for single pair mating. The eggs obtained from single pair were reared to get first generation larvae. Third instar H.armigera larvae from (1st generation) F, with an average weight of 30 mg ± 0.011 S.E. of Mahaboobnagar, Raichur and Nagpur strains were subjected separately to different concentrations of the test insecticides. The survivals at  $LD_{50}$  concentration in each test insecticide at F<sub>1</sub> (1st generation) were further used for resistance studies.

#### 2.1.4. Bioassay

Bioassay was done by topical application method using Hamilton micro applicator to evaluate the toxicity of all the test insecticides (FAO, 1971).

# 2.1.4.1. Topical application method

Initially 1.0% stock solution of the test insecticide was prepared from the formulated products by dissolving the required quantities after accurate weighment in double distilled water. The stock solution thus prepared was preserved in refrigerator for further use. Individual working concentrations for each of the test insecticides were prepared from the 1.0% stock solution through serial dilution technique using double distilled water as solvent. One micro litre of the respective insecticidal solution was applied on the dorsum of second thoracic segment by micro applicator. Three replications were maintained for each insecticidal concentration with 10 larvae in each replication.

# 2.1.5. Data collection

Mortality of the larvae was recorded at 24, 48 and 72 hours after treatment. The mortality at 72 hours after treatment was considered as end point for the assessment of toxicity of test insecticides as reported by Fisk and Wright (1992). Thus, concentrations of wide range initially and narrow range subsequently were tested so as to get mortality data in the range of 5-90 %. The moribund larvae also were considered as dead while recording the mortality data. The amount of insecticide present in one micro litre of test concentration was calculated and expressed as  $(LD_{50})$  dose in  $\mu g \mu l^{-1}$ .

# 2.2. Assessment of the degree of resistance acquired by H. armigera to the test insecticides

The mortality data of third instar H. armigera larvae of

Mahaboobnagar, Raichur and Nagpur populations of all the test insecticides were subjected to probit analysis (Finney, 1971) using POLO-PC software (Ross, 1987) to calculate LD<sub>so</sub>,  $LD_{qn}$ , Heterogeneity ( $\chi^2$ ), intercept (a), slope of the regression line (b), regression equation and fudicial limits. The degree of resistance acquired by *H. armigera* was calculated by dividing the higher LD<sub>50</sub> value of a strain with the lower LD<sub>50</sub> value of another strain among the three populations for each test insecticide and thus the relative degree of resistance was assessed (Resistance factor =  $LD_{50}$  of the resistant strain  $\div LD_{50}$ of the susceptible strain).

The degree of resistance acquired by all the three strains was also calculated by comparing the present data with the available baseline data at LD<sub>50</sub> and LD<sub>90</sub> levels. The degree of resistance to methomyl and cypermethrin were calculated by using the baseline data of Nagpur susceptible strain (Kranthi, 2005) (Table 2).

Resistance factor =  $LD_{50}$  of the  $F_1$  resistant strain ÷  $LD_{50}$  of the Nagpur susceptible strain

Table 2: Particulars of base line data used to calculate the degree of insecticide resistance in the larvae of H.armigera

Name of the	Name of	LD50 µg	LD90 μg	Refer-
insecticide	strain	larva <sup>-1</sup>	larva <sup>-1</sup>	ence
Cyperme- thrin	Nagpur sus- ceptible	0.007	0.028	Kranthi, 2005
Methomyl	Nagpur sus- ceptible	0.030	0.165	Kranthi, 2005

### 3. Results and Discussion

The larvae of H.armigera collected from Mahaboobnagar, Raichr and Nagpur selected for cypermethrin and methomyl insecticides at F<sub>1</sub> generation and they were reared to F<sub>2</sub> by single pair mating further proceeded to F<sub>3</sub> by the same single pair method, larvae subjected to these insecticides of different doses at F<sub>2</sub> and F<sub>3</sub>, the results were discussed here under.

The H. armigera larvae of the Mahaboobnagar district recorded a LD<sub>50</sub> of 29.125 μg larva<sup>-1</sup> which rose sharply to 59.609  $\mu$ g larva<sup>-1</sup> at LD<sub>90</sub> for cypermethrin at F<sub>1</sub>, which later still increased to 32.123 and 37.109 µg larva<sup>-1</sup> at LD<sub>50</sub> and LD<sub>90</sub> respectively at F, Resistance increased to 35.115 and 42.124 at LD<sub>50</sub> and LD<sub>90</sub> respectively at F<sub>3</sub> (Table 3). The LD<sub>50</sub> and LD<sub>90</sub> values of cypermethrin for Raichur population of Harmigera was 32.481 and 38.172 μg larva<sup>-1</sup>, respectively at F<sub>1</sub>, which were rosed to 35.467 and 41.875  $\mu$ g larva<sup>-1</sup> at LD<sub>50</sub> and LD<sub>90</sub>, respectively at F<sub>2</sub>, further they were increased to 38.906 and 74.202 at LD<sub>so</sub> and LD<sub>oo</sub>, respectively at F<sub>3</sub> (Table 4) Toxicity of cypermethrin to Nagpur population of H. armigera showed that the  $LD_{50}$  and  $LD_{90}$  values were 20.069 and 54.708 µg larva<sup>-1</sup>, respectively at F<sub>1</sub>, further they were sharply increased to 23.383 and 32.233  $\mu g$  larva<sup>-1</sup> at LD<sub>50</sub> and LD<sub>90</sub>, respectively at F<sub>2</sub>, later they were increased to 25.054 and 32.224 μg larva<sup>-1</sup> at LD<sub>50</sub> and LD<sub>90</sub>, respectively (Table 5) at F<sub>3</sub>

Among the three populations of *H. armigera*, the population of Raichur has developed 1.115 and 0.640 fold relative resistance at  $LD_{50}$  and  $LD_{90}$ , respectively as compared with the Mahaboobnagar population. The same Raichur population has developed still more higher levels of relative

Table	Table 3: Resistance pattern in Mahaboobnagar population of H. armigera									
SI. No.	Strain	Genera- tion	LD50 µg larva <sup>-1</sup> (95% FL)	LD <sub>90</sub> μg larva <sup>-1</sup> (95% FL)	Slope±S.E (b)	Heteroge- neity (χ²)	Regression equation			
1.	Cypermethrin	F <sub>1</sub>	29.125 (23.867 – 33.325)	59.609 (51.126-76.562)	4.120± 0.669	0.809	Y=-1.033+4.120 X			
2.	Cyper-Cyper	F <sub>2</sub>	32.123 (12.508 – 35.456)	37.109 (33.641 –97.573)	20.452± 9.654	0.777	Y=-25.818+20.452 X			
3.	Cyper-Cyper- Cyper	<b>F</b> <sub>3</sub>	35.115 (28.231 – 38.382)	42.124 (38.525–55.876)	16.213± 5.669	0.828	Y=-20.058+16.213 X			
4.	Methomyl	F <sub>1</sub>	3.651 (2.693 – 4.460)	10.287 (8.259-14.714)	2.849± 0.471	3.333	Y=3.398+2.849 X			
5.	Metho-Metho	F <sub>2</sub>	3.872 (0.569 – 4.824)	5.829 (4.667–29.441)	7.215± 3.274	0.190	Y=0.758 + 7.215 X			
6.	Metho-Metho- Metho	$F_3$	3.934 (3.285 – 4.261)	4.668 (4.305–5.971)	17.250± 5.818	0.658	Y=-5.261+17.250 X			

Table	Table 4: Resistance pattern in Raichur population of <i>H. armigera</i>									
SI. No.	Strain	Genera- tion	LD <sub>50</sub> μg larva <sup>-1</sup> (95% FL)	LD <sub>90</sub> μg larva <sup>-1</sup> (95% FL)	Slope±S.E (b)	Heteroge- neity (χ²)	Regression equation			
1.	Cypermethrin	F <sub>1</sub>	32.481 (15.858 – 35.904)	38.172 (34.615 – 87.453)	18.279± 8.376	1.091	Y=-22.630+18.279 X			
2.	Cyper-Cyper	F <sub>2</sub>	35.467 (23.520 – 38.971)	41.875 (38.106 – 62.869)	17.767± 7.332	0.611	Y=-22.537+17.767 X			
3.	Cyper-Cyper- Cyper	<b>F</b> <sub>3</sub>	38.906 (32.472 – 43.748)	74.202 (64.183-96.176)	4.571± 0.811	0.911	Y=-2.267+4.571 X			
4.	Methomyl	$F_{1}$	3.630 (2.719 – 4.408)	10.417 (8.357-14.905)	2.799± 0.450	3.321	Y=3.433+2.799 X			
5.	Metho-Metho	F <sub>2</sub>	3.659 (2.259 – 4.365)	5.471 (4.566 – 10.348)	7.337± 2.611	1.081	Y=0.866 + 7.337 X			
6.	Metho-Metho- Metho	$F_3$	3.851 (3.058 – 4.208)	4.708 (4.297 – 6.485)	14.684± 5.189	0.891	Y=-3.598 + 14.684 X			

resistance by 1.168 and 0.698 fold when compared with the Nagpur population at  ${\rm LD_{50}}$  and  ${\rm LD_{90}}$ , respectively, while Mahaboobnagar population recorded 1.451 and 1.090 fold resistance at  $LD_{50}$  and  $LD_{90}$ , respectively in comparison with Nagpur. In comparison with base line data of Nagpur susceptible population reported by Kranthi (2005) the three populations of Raichur, Mahaboobnagar and Nagpur acquired 4640.143, 4160.714, 2867.000 folds at LD<sub>50</sub> and 1363.286; 2128.893 and 1953.857 fold resistance to cypermethrin at LD<sub>90</sub> levels, respectively (Table 6).

The H. armigera larvae of the Mahaboobnagar district recorded a LD<sub>so</sub> of 3.651 µg larva<sup>-1</sup> which rose sharply to 10.287 μg larva<sup>-1</sup> at LD<sub>90</sub> for methomyl at F<sub>1</sub>. Further the values rosed to 3.872 and 5.829  $\mu g$  larva<sup>-1</sup> at LD<sub>50</sub> and LD<sub>90</sub>, respectively at  $F_2$ . Interestingly at  $F_3$  the resistance recoded was 3.934 and 4.668 at  $LD_{50}$  and  $LD_{90}$ , respectively at  $F_3$  (Table 3). The  $LD_{50}$ and  $LD_{on}$  values of methomyl for Raichur population of Harmigera was 3.630 and 10.417 μg larva<sup>-1</sup>, respectively at F<sub>1</sub>.

Table	Table 5 : Resistance pattern in Nagpur population of <i>H. armigera</i>									
SI. No.	Strain	Genera- tion	LD <sub>50</sub> μg larva <sup>-1</sup> (95% FL)	LD <sub>90</sub> μg larva <sup>-1</sup> (95% FL)	Slope±S.E (b)	Heteroge- neity (χ²)	Regression equation			
1.	Cypermethrin	F <sub>1</sub>	20.069 (15.312–24.138)	54.708 (44.089–78.011)	2.943± 0.477	2.602	Y=1.167+2.943 X			
2.	Cyper-Cyper	F <sub>2</sub>	23.383 (16.024–26.927)	32.233 (27.885–53.609)	9.193± 3.261	1.004	Y=-7.585+9.193 X			
3.	Cyper-Cyper- Cyper	F <sub>3</sub>	25.054 (18.153–28.330)	32.224 (28.485–47.966)	11.725± 4.165	0.931	Y=-11.401+11.725 X			
4.	Methomyl	$F_{1}$	2.652 (2.051–3.174)	7.214 (5.583–11.827)	2.949± 0.555	1.675	Y=3.751+2.949 X			
5.	Metho-Metho	F <sub>2</sub>	2.844 (2.083–3.199)	3.716 (3.292–5.682)	11.032± 3.906	0.953	Y=-0.007 + 11.032 X			
6.	Metho-Metho- Metho	F <sub>3</sub>	2.944 (2.607–3.088)	3.279 (3.122–3.900)	27.373± 9.661	0.805	Y=-7.836+27.373 X			

Table 6: Relative degree of resistance among the three populations of <i>H. armigera</i> to cypermethrin at F <sub>1</sub>									
Population	LD <sub>50</sub> μg	LD <sub>90</sub> μg	Resistanc	ce factor in	comparis	Resistance factor in comparison with Baseline Data			
	larva <sup>-1</sup>	larva <sup>-1</sup>	Mahabo populatio	Ū	0, 1,				
			at LD <sub>50</sub>	at LD <sub>90</sub>	at LD <sub>50</sub>	at LD <sub>90</sub>	at LD <sub>50</sub>	at LD <sub>90</sub>	
Raichur	32.481	38.172	1.115	0.640	1.618	0.698	4640.143	1363.286	
Mahaboobnagar	29.125	59.609	-	-	1.451	1.090	4160.714	2128.893	
Nagpur	20.069	54.708	-	-	-	-	2867.000	1953.857	
Baseline data (Kranthi, 2005)	0.007	0.028	-	-	-	-	-	-	

Further the values rosed to 3.659 and 5.471  $\mu g$  larva  $^{\!-1}$  at  $LD_{_{50}}$ and  $LD_{90}$ , respectively at  $F_2$ . Interestingly at  $F_3$  the resistance recoded was 3.851 and 4.708 at  $LD_{50}$  and  $LD_{90}$ , respectively at F<sub>3</sub> (Table 4). Toxicity of methomyl to Nagpur population of  $\it H. \ armigera \ showed \ that \ the \ LD_{50} \ and \ LD_{90} \ values \ were \ 2.652$ and 7.214%, respectively at F<sub>1</sub>. Further the values rosed to 2.844 and 3.716  $\mu g$  larva  $^1$  at  $L\bar{D}_{_{50}}$  and  $LD_{_{90}}$  respectively at F  $_2$ Interestingly at F<sub>3</sub> the resistance recoded was 2.944 and 3.279 at  $LD_{50}$  and  $LD_{90}$ , respectively at  $F_3$  (Table 5).

Amongst the three populations of H. armigera, the population of Mahaboobnagar has developed 1.006 and 0.988 fold relative resistance at LD<sub>50</sub> and LD<sub>90</sub>, respectively as compared with the Raichur population. The same Mahaboobnagar population has developed still more higher levels of relative resistance by 1.377 and 1.426 fold when compared with the Nagpur population at  $LD_{50}$  and  $LD_{90}$ , respectively, while Raichur population recorded 1.369 and 1.444 fold resistance at LD<sub>50</sub> and LD<sub>90</sub>, respectively in comparison with Nagpur. In comparison with base line data of Nagpur susceptible population reported by Kranthi (2005) the three populations of Mahaboobnagar, Raichur and Nagpur acquired 121.700, 121.000, 88.400 folds at LD<sub>so</sub> and 62.345, 63.133, 43.721 folds resistance to methomyl at LD<sub>90</sub> levels, respectively (Table 7).

The present results of cypermethrin resistance are in

Table 7: Relative degree of resistance among the three populations of <i>H. armigera</i> to methomyl at F <sub>1</sub>									
Population	$LD_{50} \mu g$	$LD_{90}\mu g$	Resistanc	ce factor ir	Resistance factor in com- parison with Baseline Data				
	larva <sup>-1</sup>	larva <sup>-1</sup>	Mahaboobnagar population(folds)				Nagpur popula- tion (folds)		
			at LD <sub>50</sub>	at LD <sub>90</sub>	at LD <sub>50</sub>	at LD <sub>90</sub>	at LD <sub>50</sub>	at LD <sub>90</sub>	
Raichur	3.651	10.287	1.006	0.988	1.377	1.426	121.700	62.345	
Mahaboobnagar	3.630	10.417	-	-	1.369	1.444	121.000	63.133	
Nagpur	2.652	7.214	-	-	-	-	88.400	43.721	
Baseline data (Kranthi, 2005)	0.03	0.165	-	-	_	-	-	_	

approximity with (Kranthi et al., 2001) who reported >1000 folds resistance to cypermethrin in four strains of *H. armigera* of Central India. Singh and Mahal (2005) reported that 2087 fold resistance to cypermethrin. Rao et al., (2005) reported that the synthetic pyrethroids showed high resistance frequencies (>80%) with a  $LD_{50}$  value of 8.51  $\mu g$  larva<sup>-1</sup> and it had developed highest resistance (946 folds) against cypermethrin. Chaturvedi (2007) was reported that the H. armigera exhibited widespread resistance (RF=48-919) to cypermethrin. Suryawanshi et al. (2008) revealed that the synthetic pyrethroid cypermethrin showed high resistance frequencies (84-88%) with a LD<sub>so</sub> values of 1.399 g larva<sup>-1</sup> and it had developed higher resistance (279.80 fold) against cypermethrin. Nimbalkar et al. (2009) were reported that the LD<sub>so</sub> value for cypermethrin was highest, during October (1.459 mg larva<sup>-1</sup> in Aurangabad) and lowest during August in Parbhani (0.157 µg larva-1) during 2008, which might be due to continuous use of cypermethrin to manage this pest. Achaleke et al. (2009) indicated high resistance to cypermethrin (RF=67-1771) among H. armigera field populations and laboratoryselected strains. From the present findings the resistance ratio appears to be high because of the comparison with most susceptible strain as base line (LD $_{50}$ =0.007  $\mu g$  larva $^{\text{-}1}$ ), but in reality it is indicated that there is decrease in the levels of resistance to cypermethrin in H. armigera compared to latest reports of Suryawanshi et al. (2008); Nimbalkar et al. (2009) which may be probably due to decreased selection pressure with significant decrease in the use of cypermethrin and increasing the area under Bt cotton.

The present results of methomyl resistance are in approximity with Jhansi et al., 2004 according to whom, during early part of the season on cotton the per cent resistance was found almost constant (31.34 to 64.00) and 6-30 folds of resistance to methomyl from Guntur region. Resistance level against methomyl in Khandwa district of Madhya Pradesh was found quite low (2.50%) in the first week of October and then varied from 2.50% to 18.06% during the remaining period of the month. However, the highest resistance level (44.52 %) was recorded during the third week of November (Choudhary et al., 2004). From the present investigations it is evident that there is decrease in the levels of resistance to methomyl in H. armigera compared to latest reports of Suryawanshi et al. (2008) which may be probably due to significant decrease in the use of methomyl in managing the pest due to reduced bollworm incidence and due to increased area under Bt cotton.

#### 4. Conclusion

The results obtained during the present study revealed that the continuous application of same insecticide across the generations increases the resistance from F<sub>1</sub> to F<sub>2</sub>. Alternating the new chemistries with old conventional chemicals results in no cross resistance development as it was observed for almost all populations.

#### 5. Further Research

Insecticide sequences like Cypermethrin-Cypermethrin-Cypermethrin, Methomyl-Methomyl, Cypermethrin-Methomyl-Methomyl, showed the CRR equal and above 1.0 showing future cross resistance threat and the insecticides like, Indoxacarb, Spinosad rotations may prove to be the better combinations for further study. Future studies on insecticide sequences with technical grade insecticides may give more conclusive results.

Molecular characterization of insecticide resistant *H.armigera* may through light on the nucleotide base level differences of different locations of the resistant individuals through which the resistance is governed.

#### 6. References

Achaleke, J., Martin, T., Ghogomu, R.T., Vaissayre, M., Brevault, T., 2009. Esterase mediated resistance to pyrethroids in field populations of Helicoverpa armigera (Lepidoptera: Noctuidae) from Central Africa. Pest Management Science 65(10), 1147-1154.

Armes, N.J., Jadhav, D.R., Desouza, K.R., 1996. A survey of insecticide resistance in Helicoverpa armigera in the Indian subcontinent. Bulletin of Entomological Research 86, 499-514.

Chaturvedi, I., 2007. Status of insecticide resistance in the cotton bollworm, Helicoverpa armigera (Hubner). Journal of Central European Agriculture 8(2), 171–182.

Choudhary, R.K., Yadav, A.S., Shrivastava, V.K., Tomar, S.P.S., Patel, Y., 2004. Studies on insecticide resistance in Helicoverpa armigera (Hubner) on cotton in Madhya Pradesh. In: International Symposium on Strategies for Sustainable Cotton Production – A global vision 3. Crop Protection, University of Agricultural Sciences, Dharwad, Karnataka, India, 23-25 November.

FAO., 1971. Recommended methods for the detection and measurement of pest resistance to pesticides. Tentative method for larvae of Egyptian cotton leafworm (Spodoptera littoralis Biosd.) F.A.O method No.8 Food and Agricultural Organization. Plant Protection Bulletin 19, 32-35.

Finney, D.J., 1971. Probit analysis, Cambridge University, London, 333.

Fisk, T., Wright, D.J., 1992. Speed of action and toxicity of acylurea insect growth regulators against Spodoptera exempta (Walk.) and Spodoptera littoralis (Biosd.) larvae: effect of inter moult age. Pesticide Science 35, 331–337.

Jhansi, K., Radhika, P., Subbaratnam, G.V., 2004. Insecticide resistance in insect pests of cotton in Andhra Pradesh. In: International Symposium on Strategies for Sustainable Cotton Production-A global vision 3. Crop Protection, University of Agricultural Sciences, Dharwad, Karnataka, India, 23-25 November.

Kranthi, K.R., 2005. Insecticide resistance monitoring,

- mechanisms and management manual. Central Institute for Cotton Research, Nagpur, 80-94.
- Kranthi, K.R., Armes, N.J., Rao, N.G.V., Raj, S., Sundaramurthy, V.T., 1997. Seasonal dynamics of metabolic mechanisms mediating pyrethroid resistance in Helicoverpa armigera in central India. Pesticide Science 50, 91-98.
- Kranthi, K.R., Jadhav, D.R., Wanjari, R.R., Ali, S.S., Russell, D., 2001. Carbamate and organophosphate resistance in cotton pests in India 1995 to 1999. Bulletin of Entomological Research 91(1), 37-46.
- Manjunath, T.M., 1990. Controlling cotton pests. Deccan Herald, January 4,1990 Science and society, 1–3.
- Martin, T., Ochou, G.O., Vaissayre, M., Fournier, D., 2003. Monitoring of insecticide resistance in *Helicoverpa* armigera (Hubner) from 1998 to 2002 in Cote d'Ivoire, West Africa. Resistant Pest Management Newsletter 12(2), 51-55.
- Nimbalkar, R.K., Shinde, S.S., Tawar, D.S., Muley, S.P., 2009. Response of cotton bollworm Helicoverpa armigera (Hubner) (Lepidoptera: noctuidae) to different insecticides in Maharashtra, India. World Journal of Agricultural Sciences 5(2), 250-255.
- Singh, N., Mahal, M.S., 2005. Acute toxicity of different insecticides against Helicoverpa armigera (Hubner) in Punjab. Pesticide Research Journal 17(2),52–54.

- Ramasubramanian, T., Regupathy, A., 2004. Magnitude and mechanism of insecticide resistance in Helicoverpa armigera (Hub.) population of Tamil Nadu, India. Asian Journal of Plant Sciences 3(1), 94-100.
- Rao, G.M.V.P., Rao, N.H., Raju, K., 2005. Insecticide resistance in field population of American bollworm, Helicoverpa armigera (Hub.) (Lepidoptera: Noctuidae). Resistant Pest Management Newsletter 15(1), 15–17.
- Reed, W., Pawar, C.S., 1982. Heliothis, a global problem. In: Proceedings of International Workshop on Heliothis Management, ICRISAT, Patancheru, Andhra Pradesh, India, 9–14.
- Ross, G.J.S., 1987. Maximum likely hood program. The numerical Algorithms Group. Rothamsted Experimental Station, Harpenden, UK.
- Suryawanshi, D.S., Bhede, B.V., Bhosale, S.V., More, D.G., 2008. Insecticide resistance in field population of American bollworm, Helicoverpa armigera (Hub.) (Lepidoptera: Noctuidae). Indian Journal of Entomology 70(1), 44-46.
- Zhou, X., Fartor, O., Applebaum, S.W., Coll, M., 2000. Population structure of the pestiferous moth Helicoverpa armigera in the Eastern Mediterranean using RAPD analysis. Heredity 85, 251-256.